4/27/16

- Continuation of book 4186

- Removed cocktail biomarker 2016 frogs from freeze-dryer
- Extracted Day 1 frogs

  - Chopped up frogs
  - Added 7.5mL of MeOH → vortex + sonicate for 30min w/ice bath
  - Centrifuge for 20min @ 3250rpm
  - Transfer supernatant to a 20mL scint vial
  - Add 5mL of MeOH → vortex + sonicate for 30min w/ice bath
  - Centrifuge for 20min @ 3250rpm
  - Transfer supernatant to corresponding 20mL scint vial + blow down to 1mL under N_2 gas
  - Add 10mL MQ → vortex
  - Add 3mL MTBE → vortex
  - Add scoop of Na_2SO_4 → vortex
  - Transfer top organic layer to a pre-labeled 2mL centrifuge tube
  - Centrifuge at 14000 rpm for 15min.
  - Transfer 1mL aliquot to a prelabeled GC vial + placed in freezer until analyzed.

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	4/27/16

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Book No. 4 TITLE Bertha LC/MS/MS - Bees method

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4/28/16

- Finalized method for pollen, honey + bee samples.
- Used a hypersil Gold 50 x 2.1 1.7um column by Thermo with a guard cartridge

- Method: Pollen and honey — 7030

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*************************************
   ProStar 230 Inert SDM
*************************************
Module Address: 01

          Solvent A            : H20 w/ 0.05 % FA and 2mM AmAc
          Solvent B            : MeOH w/ 0.05 % FA and 2mM AmAc
          Solvent C            : ACN + 0.1% FA
          Maximum Pressure     : 400 atm
          Minimum Pressure     :   0 atm
   0.00   Relays               : ------
   0.00   Flow Rate            : 0.30   ml/min
   0.00   Composition          :  95% A,   5% B,   0% C
   0.00   Equilibration Time   : 10.00  min
   2.00   Relays               : ------
   2.00   Flow Rate            : 0.30   ml/min
   2.00   Composition          :  95% A,   5% B,   0% C
   9.00   Relays               : ------
   9.00   Flow Rate            : 0.30   ml/min
   9.00   Composition          :   1% A,  99% B,   0% C
  12.00   Relays               : ------
  12.00   Flow Rate            : 0.30   ml/min
  12.00   Composition          :   1% A,  99% B,   0% C
  12.10   Relays               : ------
  12.10   Flow Rate            : 0.30   ml/min
  12.10   Composition          :  95% A,   5% B,   0% C
  28.00   Relays               : ------
  28.00   Flow Rate            : 0.30   ml/min
  28.00   Composition          :  95% A,   5% B,   0% C
  28.00   Method End Action    : Equilibrate at End
*************************************
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DAG

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*****************************************************
Acquisition Segment Information
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Method Time: 0.02-28.08, Centroid, ESI
  Seg 1, Time: 0.02-9.82, Scan Functions: 8
    1. 203.0>113.0 [-12.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    2. 203.0>129.0 [-9.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    3. 271.0>126.0 [-16.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    4. 271.0>196.0 [-16.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    5. 292.0>181.0 [-22.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    6. 292.0>211.0 [-10.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    7. 295.0>184.0 [-16.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    8. 295.0>214.0 [-12.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
  Seg 2, Time: 9.83-28.08, Scan Functions: 8
    1. 223.0>90.0 [-28.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    2. 223.0>126.0 [-18.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    3. 250.0>132.0 [-12.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    4. 250.0>169.0 [-11.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    5. 253.0>90.0 [-29.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    6. 253.0>126.0 [-17.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    7. 256.0>175.0 [-19.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500
    8. 256.0>209.0 [-15.0V] ESI;CID 80.0;Det 1220;WI Q1 1.00;Q3 1.00;ST 0.500

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203 - Dinotefuran
271 - Nitenpyram
292 - Thiamethoxam
295 - Thiamethoxam-D
223 - Acetamiprid
250 - Clothianidin
253 - thiacloprid
256 - imidacloprid

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Vial	Sample	Vol	Method	Data File	Instrument
F1	BLANK	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F2	0.0039	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F3	0.125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F4	0.0625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F5	0.0078	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	0.03125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F7	0.0156	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F8	0.25	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E1	6-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E2	3-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E3	10-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E4	8-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E5	7-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E6	2-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E7	5-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E8	9-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D1	4-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D2	1-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D3	18 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D4	37 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D5	67 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D6	75 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D7	89 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D8	33 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C1	55 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C2	49 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C3	87 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C4	73 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C5	52 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C6	32 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C7	30 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C8	14 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B1	35 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B2	58 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B3	64 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B4	83 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B5	78 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B6	24 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B7	44 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B8	22 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A1	28 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A2	80 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A3	16 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A4	69 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha

DAG

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Vial	Sample	Vol	Method	Data File	Instrument
A5	76 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A6	72 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A7	41 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A8	85 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F2	0.0039	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F3	0.125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F4	0.0625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F5	0.0078	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	0.03125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F7	0.0156	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F8	0.25	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E1	34 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E2	93 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E3	42 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E4	71 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E5	91 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E6	13 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E7	65 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E8	62 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D1	53 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D2	84 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D3	47 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D4	36 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D5	29 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D6	38 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D7	82 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D8	54 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C1	57 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C2	51 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C3	56 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C4	25 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C5	17 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C6	60 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C7	45 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C8	21 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B1	77 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B2	39 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B3	27 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B4	59 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B5	74 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B6	40 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha

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Vial	Sample	Vol	Method	Data File	Instrument
B7	46 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B8	26 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A1	66 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A2	43 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A3	94 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A4	81 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A5	70 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A6	11-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A7	23 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A8	79 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F2	0.0039	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F3	0.125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F4	0.0625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F5	0.0078	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	0.03125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F7	0.015625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F8	0.25	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E1	20 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E2	19 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E3	90 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E4	92 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E5	12-Apr	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E6	15 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E7	31 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E8	48 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D1	61 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D2	63 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D3	86 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D4	88 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D5	68 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D6	50 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D7	7-198 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D8	7-195 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C1	7-182 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C2	7-185 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C3	7-188 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C4	7-191 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C5	7-186 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C6	7-200 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C7	7-196 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C8	7-187 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha

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Witnessed & Understood by me,

Date

Invented by:

Date 4/28/16

Recorded by:

Project No. _____

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Vial	Sample	Vol	Method	Data File	Instrument
B1	7-175 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B2	7-162 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B3	7-158 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B4	7-178 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B5	7-163 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B6	7-166 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B7	7-156 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B8	7-161 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A1	7-152 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A2	7-164 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A3	7-170 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A4	7-171 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A5	7-160 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A6	7-173 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A7	7-172 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A8	7-155 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F2	0.0039	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F3	0.125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F4	0.0625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F5	0.0078	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	0.03125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F7	0.015625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F8	0.25	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E1	7-181 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E2	7-154 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E3	7-176 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E4	7-168 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E5	6-133 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E6	6-138 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E7	6-132 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E8	6-142 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D1	6-139 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D2	6-141 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D3	6-151 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D4	6-136 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D5	6-145 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D6	6-144 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D7	6-147 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D8	6-149 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C1	6-135 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C2	6-137 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C3	6-146 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C4	6-143 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha

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Witnessed & Understood by me, | Date | Invented by: | Date 4/28/16

Recorded by:

TITLE Pollen Analyzed on Bertha

Project No.

Book No. 4

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4/28/16 - 5/4/16

Vial	Sample	Vol	Method	Data File	Instrument
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C5	6-150 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C6	6-134 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C7	6-140 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C8	6-148 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B1	8-212 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B2	8-205 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B3	8-220 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B4	8-217 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B5	8-215 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B6	8-210 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B7	8-226 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B8	8-216 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A1	8-222 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A2	8-230 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A3	8-211 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A4	8-231 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A5	8-229 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A6	8-228 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A7	8-227 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A8	8-224 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F2	0.0039	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F3	0.125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F4	0.0625	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F5	0.0078	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	0.03125	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F7	0.0156	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F8	0.25	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E1	8-225 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E2	8-221 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E3	8-209 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E4	8-208 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E5	8-223 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E6	8-218 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E7	8-214 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
E8	8-207 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D1	8-213 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D2	8-204 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D3	8-219 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D4	8-206 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D5	7-190 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D6	7-199 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D7	7-197 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
D8	7-183 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	4/28/16

Project No.
Book No. 4
TITLE Pollen Analyzed on Bertha

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4/28/16 - 5/4/16

Vial	Sample	Vol	Method	Data File	Instrument
C1	7-193 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C2	7-192 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C3	7-189 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C4	7-203 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C5	7-202 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C6	7-201 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C7	7-194 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
C8	7-184 p*2	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B1	7-167 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B2	7-180 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B3	7-165 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B4	7-157 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B5	7-169 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B6	7-177 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F6	qaqc	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B7	7-179 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
B8	7-153 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A1	7-174 p	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
A2	7-159	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha
F1	blank	20	Pollen and honey_7030	DAG/Neonic/Pollen_2 use	Triple quad-Bertha

DAG

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Witnessed & Understood by me, | Date | Invented by: | Date 4/28/16
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TITLE Nurse Bees & Triclosan extraction

Project No. _____

Book No. 4

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4/29/16

- Crushed up 12 nurse bees
  - ~2 grams of nurse bees were in a ziploc bag
  - Spiked with 2uL of 1000ppm thiamethoxam D3
  - Added 2 scoops (~2g) of diatamaceous earth
  - Crushed up with a rolling pin until fine grain
  - Poured into a pre-labeled 15mL centrifuge tube
  - Added 1-2mL of EtOH to bag - hand massage
  - Poured into the same 15mL centrifuge tube
  - Placed on speedvac overnight

5/2/16

- Finished blowing down Keri's sediment samples for september until ~2mL of MeOH - Triclosan from book 4/86

5/3/16

- Finished blowing down Keri's sediment samples for october until ~2mL of MeOH - Triclosan

5/4/16

- Finished blowing down Keri's sediment samples for november until ~2mL of MeOH - Triclosan

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	5/4/16

TITLE Triclosan standard

Project No.

Book No. 4

From Page No.

5/3/16 - 5/9/16

Vial	Sample	Method	Data File	Data Line	Instrument
98	BIF 10 4 S	Pest_Biomarker_SIM	DAG/Frog Biomarkers	160503256	GCMS-s.q.
99	TM MAX 2 S	Pest_Biomarker_SIM	DAG/Frog Biomarkers	160503257	GCMS-s.q.
100	TM MAX 6 S	Pest_Biomarker_SIM	DAG/Frog Biomarkers	160503258	GCMS-s.q.

DAG

5/6/16

- created new standards for triclosan

5/6/16

Triclosan

z = 2uL
X = 1mL

1000ppm (z) = 2mL (1ppm)

1ppm (x) = 2mL (0.5ppm)
0.5ppm (x) = 2mL (0.25ppm)
0.25ppm (x) = 2mL (0.125ppm)
0.125ppm (x) = 2mL (0.0625ppm)
0.0625ppm (x) = 2mL (0.03125ppm)
0.03125ppm (x) = 2mL (0.015625ppm)
0.015625ppm (x) = 2mL (0.0078 ppm)
0.0078 ppm (x) = 2mL (0.0039ppm)
0.0039ppm (x) = 2mL (0.0019ppm)

DAG

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	5/6/16

Project No. _____
Book No. 4 TITLE Triclosan, Pollen LOD

From Page No. ____

5/9/16

- Transferred all of the Triclosan sediment samples to GC vials and blow down under nitrogen along with 3 MeOH rinses

- Once dry samples were reconstituted in 1mL 30% MeOH + analyzed on Bertha LC-triple quad.

- For Pollen Samples
  - the standard curve was created using all standards + qaqc standards
  - where the intercept was set at 0.

- ~~St~~ Detection limit
  - quantify all the blank samples
  - $\frac{3 \cdot \text{stdev (blanks)}}{\text{slope}}$ Detection limit but for Ø values can do Ø or DL/2.

- Extracted remaining livers from cocktail Biomarker 2016 study following extraction on pg. 154 of book 4/83

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	5/9/16

TITLE Triclosan Analyzed on Bertha

Project No.

Book No. 4

From Page No.

5/10/16 - 5/14/16

Vial	Sample	Vol	Method	Data File	Instrument
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F2	0.0039	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F3	0.0078	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F4	0.015625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	0.0625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F7	0.125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F8	0.25	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E1	0.5	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E2	1	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E3	FR1B 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E4	DB3B 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E5	DB4C 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E6	DB4B 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E7	DB2C 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E8	FR2C 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D1	FR1C 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D2	DB3A 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D3	DB2A 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D4	DB4C 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D5	FR1B 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D6	FR2B 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D7	DB3C 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D8	FR2A 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C1	DB3B 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C2	DB4B 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C3	DB1B 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C4	DB4A 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C5	FR2C 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C6	DB3A 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C7	DB2B 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C8	FR1A 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B1	DB1C 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B2	DB4A 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B3	FR1A 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B4	DB3C 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B5	FR1C 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B6	FR2A 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B7	DB1A 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B8	DB1B 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A1	DB2A 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A2	DB1C 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A3	DB2C 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A4	DB2B 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A5	DB1A 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A6	DB1B 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha

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Date 5/10/16

Recorded by:

Project No.
Book No. 4
TITLE Triclosan Analyzed on Bertha
5/10 - 5/14/16

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Vial	Sample	Vol	Method	Data File	Instrument
A7	DB4A 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F2	0.0039	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F3	0.0078	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F4	0.015625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	0.0625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F7	0.125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F8	0.25	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E1	0.5	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E2	1	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A8	DB4C 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E3	DB2C 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E4	DB3B 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E5	FR2C 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E6	DB4A 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E7	FR2C 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E8	DB3A 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D1	FR2A 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D2	DB1C 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D3	DB3C 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D4	DB1A 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D5	FR2A 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D6	DB2A 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D7	DB4B 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D8	DB3C 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C1	FR1B 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C2	DB4C 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C3	DB3A 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C4	FR2B 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C5	FR1C 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C6	DB1C 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C7	DB3B 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C8	DB2B 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B1	FR1A 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B2	DB2C 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B3	DB2B 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B4	FR1A 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B5	DB2A 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B6	FR1B 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B7	DB1A 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B8	FR1C 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A1	FR2B 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A2	DB4B 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A3	DB1B 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A4	EFF 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A5	EFF 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha

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Recorded by:

Date 5/10/16

TITLE Triclosan Analyzed on Bertha

Project No.

Book No.

From Page No.

5/10/16 - 5/14/16

Vial	Sample	Vol	Method	Data File	Instrument
A6	EFF 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A7	EFF 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A8	INF 11W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E3	INF 9W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E4	INF 10W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E5	INF 8W	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F2	0.0039	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F3	0.0078	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F4	0.015625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	0.0625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F7	0.125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F8	0.25	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E1	0.5	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E2	1	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E6	db1b 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E7	db2c 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E8	db3b 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D1	db4a 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D2	db4b 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D3	fr2c 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D4	db1a 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D5	db1c 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D6	db2a 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D7	db1a 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D8	db2b 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C1	fr2a 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C2	db4c 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C3	db1b 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C4	db1c 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C5	db3c 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C6	db2b 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C7	db3b 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C8	db3c 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B1	db4a 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B2	db3a 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B3	fr1b 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B4	fr1a 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B5	db2a 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B6	db2c 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B7	db4b 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B8	db3a 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A1	fr1c 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A2	db3c 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A3	fr2b 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha

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Recorded by:

Project No.
Book No. 4 TITLE Triclosan
From Page No.
5/10/16 - 5/14/16

Vial	Sample	Vol	Method	Data File	Instrument
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A4	fr2b 8s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A5	fr1c 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A6	fr2a 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A7	fr2c 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A8	fr1b 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E3	fr1a 9s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F2	0.0039	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F3	0.0078	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F4	0.015625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	0.0625	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F7	0.125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F8	0.25	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E1	0.5	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E2	1	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E4	db1c 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E5	db2a 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E6	db2b 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E7	fr2c 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
E8	fr1c 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D1	db4c 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D2	fr2b 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D3	db2c 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D4	db1a 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D5	db1a 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F6	QAQC	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D6	db1b 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D7	db1b 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
D8	db3a 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C1	fr2a 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C2	fr2c 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C3	db4c 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C4	fr2a 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C5	db4a 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C6	db3b 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C7	fr1b 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
C8	db3c 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B1	db4b 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B2	fr1a 10s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B3	db2a 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B4	db1c 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B5	db4b 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B6	db3a 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B7	db2c 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
B8	fr2b 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A1	db3c 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	BLANK	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha

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Witnessed & U
Recorded by:
Date 5/10/16

TITLE Triclosan, honeybees from Nuebox

Project No.

Book No. 4

From Page No.

5/10 - 5/14/16

Vial	Sample	Vol	Method	Data File	Instrument
A2	fr1b 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A3	fr1a 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A4	fr12c 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A5	db4a 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A6	db2b 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
A7	db3b 11s	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F5	0.03125	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha
F1	blank	50 uL	Triclosan_neg	Triclosan	LC triple quad-Bertha

MB

5/13/16

- Chopped/crushed up 24 bee samples from Nuc box 2
- Samples were spiked with 2uL of 1000ppm thiamethoxam D3
- Added ~2 grams of diatomaceous earth
- Crushed up with rolling pin
- Transferred to a pre-labeled 15mL centrifuge tube
- Squirted EtOH in the ziploc bag & hand massage to get residu
- Transferred to 15 mL centrifuge tube
- Placed on Speedvac overnight

5/16/16

- Crushed up 20 more nurse bees from nuc box 3-5 using the above procedure

5/17/16

- Crushed up 20 nurse bees from nuc box 3-5 using the above procedu
- Yesterday's + today's bees went on the Speedvac overnight

5/18/16

- Crushed up remaining nurse bees from nuc box 3-5 using the above procedure.
- Placed samples on speedvac overnight

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	5/18/16

Project No. _____
Book No. 4 TITLE Nurse bees from nucbox

From Page No. ____

5/19/16

- Crushed up honeybees from dead bee traps, using procedure on page 21 of book 4206.
- Crushed up honeybees from in hive experiment using procedure on pg. 21
- Samples went on speedvac overnight

5/25/16

- Crushed up honeybees from dead bee traps (#60-80) using procedure on pg. 21 of book 4206.
- Samples after in fridge overnight

5/26/16

- Crushed up honeybees from dead bee traps #(30-60) using procedure on pg 21 of book 4206.

5/27/16

- Crushed up honeybees from ~~dead bee trap~~ & in hive ~~dead~~ experiment following procedure on pg. 21 of book 4206.

6/6/16

- Placed ~~on~~ nursebees from nuc box 3-5 on accelerated solvent extractor that is downstairs.
  - There are 12 stainless steel 10mL cells.
  - Place a 27mm fiber glass filter on the bottom frit endcap + push down with black tool so that it is in place w/out covering PEEK O-ring.
  - Added one scoop of alumina B
  - Added entire bee sample
  - Pushed/compacted everything using black tool.
  - Top the cell off with ottawa sand
  - Brush the threads clean with a brush and place the top endfrit on.

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	6/10/16

TITLE ASE method, honeybee extraction

Project No. _____

Book No. 4

From Page No. ____

6/6/16

- Use method #1:

Temperature: 100°C Heat: 5 min
Static time: 5 min min cycles: 2
Rinse volume: 60% Purge: 60s
Solvent A: Ethyl acetate (2)
Solvent B: methanol (1)

- Fill up collection trays with 60mL collection vials
  ↳ Menu → sequence editor → fill it → save at top
    ↳ menu → load method/seq → fill it → enter → start
- Once samples are done place collection vials in fridge
- Remove SS extraction cells and empty them into trash
- Clean with water + then methanol let air dry overnight

6/7/16

- Processed 12 more Nuc box nursebees on the ASE
- Sample extracts were placed on the fridge

6/8/16

- Processed 12 nuc box nursebees on the ASE

6/9/16

- Processed more nuc box nursebees on the ASE

6/10/16

- Processed 8 nucbox nursebees on the ASE
- Started to filter the samples through acrodisc LC 25mm syringe filters w/ 0.45 um PVDF membrane using a vacuum manifo
- New inserts were added precleaned w/ MeOH and a syringe between each sample.
- The filtered extracted were collected in 20mL glass test tubes along with 3 solvent rinses
- Samples were blown down under nitrogen gas

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	6/10/16

Project No.
Book No. 4 TITLE Honeybees extracted

From Page No.

6/13/16

- Processed several more nucbox nursebees on the ASE
- Blowing down samples under nitrogen
- Filtered more samples following procedure on pg. 23

6/14/16

- Processed more nucbox nursebees using the ASE from pg. 23
- Filtered more samples using procedure on pg. 23
- Blowing down more samples under nitrogen

6/15/16

- Processed more nucbox nursebees using the ASE
- Blowing down more samples under nitrogen gas
- filtered more samples using procedure on pg. 23

6/16/16

- Processed more nucbox nurse bees using the ASE
- Blowing down more samples under nitrogen gas

6/17/16

- Processed more nucbox nurse bees using the ASE
- Blowing down more samples under nitrogen gas

6/22/16

- Processed more nucbox nurse bees using the ASE
- Blowing down more samples under nitrogen gas

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	6/22/16

TITLE Honeybee extracted + triclosan

Project No.

Book No. 4

From Page No.

6/23/16 - 7/1/16

- Processed Dead bee traps (DBT) using the ASE
- Blowing down samples under nitrogen gas

7/5/16

- Processed DBT using the ASE
- Blow down samples under nitrogen gas
- Filtered samples through 0.2um filter after reconstituted with 1mL 30% MeOH and vortex ~~for~~ until well mixed.
- Transferred to 2mL GC vial + placed in fridge

7/6/16

- Processed DBT honeybee samples using the ASE
- Blow down samples under nitrogen gas

7/12/16

- Extracted Triclosan samples from April
- Preconditioned a 3cc (60mg) cartridge with 3mL MeOH 3mL M
- Sample passed through, rinsed with 5mL MQ
- Vacuum dry for 45 minutes
- Elute with 6mL MeOH into a clean test tube
- Blow down under nitrogen gas
- Transfer to a prelabeled GC vial with 3 solvent rinses (MeO
- Blow to dryness and reconstitute in 30% MeOH: MQ 1mL

7/15/16

- Extracted May + June triclosan samples following the same procedu as above.

- Effluent water from April, May + June were extracted on a 150mg 3 cartridge with 0.45um filters beforehand to remove particles
- Extraction is similar as 7/12/16

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	7/14/16

From Page No. ____

7/15/16

- Triclosan influents were extracted for April, May + June on 500mg 6 cartridges.

- Influents were extracted using the same procedure as on 7/12/16.

7/27/16

- Extracted more DBT honeybees on ASE
- Blow down samples
- Placed alachlor, DIA, DEA, ATZ, alachlor OXA, alachlor ESA, 2-4D on gcms for retention time and m/z values (single quad)

7/28/16

- Extracted honeybees on ASE - in hive
- Filtered NB honeybees
  - after drying 60mL scintillation vial down ~3mL left on vacuum box
    - replace liners with new ones + clean with MeOH
    - Place labeled test tubes in holder
    - Using 3mL syringe and 0.25um filter attach to top of vacuum box
    - Vortex sample for 10 sec + add to syringe
    - Squirt scint vial w/ MeOH for 2 rinses and add to syringe
    - squirt with MeOH + EtOAc - vortex add to syringe
    - squirt scint vial w/ EtOAc - vortex add to syringe
    - Use plunger to get all of sample through filter
    - Transfer test tube to nitrogen evaporator with water bath
    - Blow down until dry
    - Reconstitute with 10% MeOH: H_2O (1mL) vortex until well mixed
    - Centrifuge sample to get sides
    - Transfer sample to prelabeled 2mL centrifuge tube w/ 0.2um filter
    - Centrifuge for 15min @ 14000rpm
    - Transfer to prelabeled GC vial + place in fridge

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	7/28/16

TITLE Honeybee extraction, Bertha - MB method

Project No. _____

Book No. 4

From Page No. ____

7/29/16

- Run more samples (honeybees on ASE) - in hive
- Filter more honeybees + blow down

~~7/30/16~~ 8/1/16

- ~~Analyzed~~ Processed more honeybees on ASE - in hive
- Filtered + blow down more nb honeybees following procedure on 7/28/16

From 7/13/16 - 8/1/16 analyzed mixture biomarker on Bertha LC/MS/MS.

Solvent A - Water w/ 2mM Ammonium Acetate + 0.05% Formic
Solvent B - MeOH w/ 2mM Ammonium Acetate + 0.05% Formic Acid
- Looked for: MESA, MOXA, TDN, TDL, Tetraconazole, Metolachlor, Bifenthrin, P
- 40min method flow 400µl/min
- column: Eclipse XDB - C18 3.5um 3.0 x 150 mm Agilent with XDB-C18 guard cartridge

MOXA: 278 → 158 (+20V)
278 → 286 (+9 V) *

MESA: 328 → 80 (+25V)
328 → 121 (+17V) *

Metolachlor: 284 → 176 (-25V)
284 → 252 (-14V)*

TDN: 294 → 197 (-15V)
294 → 225 (-13V)*

TDL 296 → 70 (-17V)*
296 → 96 (-14V)

Tetra: 372 → 70 (-19 V)
372 → 159 (-30 V)

Per: 408 → 153 (-17 V)
408 → 183 (-22V)*

Bif: 440 → 166 (-40 V)
440 → 181 (-21V)*

- Half way thru bif disappeared so it had to be reran

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	8/1/16

Project No.
Book No. 4 TITLE MB samples analyzed + honeybee extraction
m Page No.

7/13/16 - 8/1/16

SAMPLE	VOL	METHOD	DATA FILE	INSTRUMENT
0.625 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.0078	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.00976	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.078 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.009 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
2.5 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.019 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
1.25 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.039 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
0.15625 TDL	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
BLANK	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
BLANK	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
BLANK	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha
BLANK	20 uL	Cocktail_biomarker_pos	DAG/Cocktail biomarkers 2016	LC triple quad-Bertha

DAG

8/2/16

- Processed the last batch of honeybees on the ASE
  Filtered more nb honeybees following procedure on 7/28/16
- Blow down honeybee samples

8/3/16

- Filtered more nb + DBT honeybees that were blown down following procedure on 7/28/16

Blow down more honeybee samples

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	8/3/16

TITLE MB metabolomic samples

Project No.

Book No. 4

From Page No.

8/3/16

- Mixture biomarkers
  were derivatized (cm 1-6, TDN 10, Met 10, BIF 10)
  - Remove samples from freezer
  - Made a solution of methoxyamine hydrochloride : MeOx
    ~ 60 mg in 3 mL of pyridine → 20 mg/mL

- Added 100 µL of MeOx to each sample, capped, vortex + placed in oven @ 60° for 2.5 hrs - vortex every 30 min.

- Removed from oven let cool for 10 min.

- Added 100 µL of BSTFA + 10% TCMS to each vial, vortexed + pl back in oven for 1.5 hrs. - vortex every 30 min

- Removed from oven + let cool for 10 min

- Transfer sample to an insert + place back into vial

- Ran samples on GC x GC - ToF for first 17 samples w/ liquid r

- Ran on GC-ToF with a blank every 10 samples
  - 1D metabol
  - 1D metabol
  - Full scan s/N 50

8/4/16

- Blow down + filtered more NB + DBT honeybees

8/5/16

- Derivatized next batch of mixture biomarker (cm 7-12, TB 10, TM 10, BM 10, following procedure from 8/3/16

To Page No

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	8/5/16

Project No. _____
Book No. 4 TITLE MB metabolomics + honeybee, triclos

From Page No. ____

8/8/16

- Blow down more honeybees DDT + in hive

8/11/16

- Derivatized another batch of mixture biomarker (TDN max, BIFmax, r con 13-18) following procedure from 8/3/16.
- Placed samples on TOF

8/12/16

- Derivatized last batch of mixture biomarker (con 19-24, TB max, TM max, MBma following procedure from 8/3/16
- Placed samples on TOF

8/15/16

- Blow down April sediment samples for Triclosan from Keri Ly

8/16/16

- Blow down May + June sediment samples for Triclosan from Ke Lydon

8/17/16

- Blow down remaining sediment - triclosan samples to ~1m

8/18/16

- Continue blowing down sediment - triclosan samples to ~1mL
- Run several mixture biomarker - max samples in 4D GCxGC,

To Page No

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	8/18/16

8/23/16

- Filtered May sediment samples for triclosan following procedure on 8/19/16
- Filtered bee samples following procedure on 7/28/16
- Finished reconstituting April + some May samples in 1mL 30% Me
- Placed honeybee samples from 2013 + 2014 on freeze-dried overnight

8/24/16

- Filtered more honeybee samples from procedure on 7/28/16
- Filtered remaining June sediment triclosan samples for Kai following procedure on 8/19/16
- Reconstitute April, may + June sediment triclosan with 1mL of 30% MeOH : H_2O
- Blow down + reconstitute bees samples in 10% MeOH + filter with 0.2um filter and centrifuge for 20min @ 14000rpm transfer to pre-labeled GC vial

8/25/16

- Filtered more honeybees following procedure on 7/28/16
- Blow down + reconstitute with 1mL 10% MeOH : H_2O
- Finished April sediment triclosan samples by reconstituting with 30% MeOH

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	8/25/16

Project No.
Book No. 4
TITLE extracted honeybees

From Page No.

8/26/16

- Filtered more honeybees using procedure from 7/28/16
- Blow down + reconstituted honeybees with 1mL 10% MeOH
- Reconstituted remaining May + June sediment triclosan samples with 1mL 30% MeOH

8/29/16

- Placed more honeybees on freeze-dried
- Weigh out 2grams of bee bread into 15mL centrifuge tube
- Blow down + reconstitute DBT samples following procedure from 7/

8/30/16

- Spiked all bee bread samples with 2uL of 1000ppm thiamethoxam
- Added 2mL of MQ, to each sample + vortex for 2 min until are mixed
- Centrifuge for 20 min + placed in fridge overnight
- Placed more honeybee samples on freeze-dried
- Weighed out 5g of honey into a 15mL centrifuge tube
- Blow down + reconstitute DBT + 1H samples following procedure on 7/28/16.

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Witnessed & Understood by me, | Date | Invented by: | Date 8/30/16

TITLE honeybee extraction

Project No. _____

Book No. 1

From Page No. ____

8/31/16

- Placed more nursebees on the freezer dries overnight
- Blow down + reconstitute in 1mL 10% MeOH more nursebees
- Weighed out 2 grams of honeybees into ziploc bags, recording values

9/1/16

- Weighed out 2 grams of honeybees into ziploc bags, recording values

9/12/16 - 9/16/16

- Added 2uL of 1000ppm thiamethoxam D3 to each sample
- Added 2 scoops (~2 grams) of diatomaceous earth to each sam
- Crush up everything with hands and rolling pin until fine powder
- Transfer sample to a prelabeled 15mL centrifuge tube using funn
- Add a small amount of EtOH to bag + hand massage bag to remove excess bee residue
- Transfer to 15mL centrifuge tube using funnel
- Repeat EtOH step a second time

9/20/16

- Finished crushing bees following procedure listed above.

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	9/20/16

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Honeybees 9/21/16

- Finished filtering honey bee samples including In hive bees + some honeybees from a 2016 bee kill.

- Samples were filtered following procedure from 7/28/16

- After filtering they were placed under nitrogen gas with a water bath @ 40°C to help evaporate solvent.

- After samples were blown to dryness they were reconstituted in 10% MeOH : H_2O 1mL.
  → Samples were vortexed until well mixed
  → Transfered to a 2mL centrifuge tube with a 0.2um filter
  → Centrifuged for 15min @ 14000 rpm

- After centrifugation samples were transfered to a 2mL GC vial and placed in freezer with other Samples until analyzed

9/22/16

Honeybees
- Processed several honeybee samples from 2013-2014 on ASE using method from pg. 23 6/6/16

9/23/16

Honeybees
- Processed several honeybee samples from 2013-2014 on ASE using method from pg. 23 6/6/16

- Blow down the 60mL scint vials under nitrogen until ~3mL were left.

To Page No.

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	9/23/16

TITLE honeybee extraction

Project No. _____

Book No. 4

From Page No. ____

Honeybees 9/26/16

- Processed several honeybee samples from 2013-2014 on A
  using method from pg. 23 6/16/16
- Blow down samples under nitrogen

Honeybees 9/27/16

- Processed several more honeybee samples from 2013-2
  on ASE downstairs using method from 6/6/13 pg. 23

Honeybees 9/28/16

- Finished processing several (fast) honeybees from 2013-2
  on ASE downstairs using method from 6/6/13 pg.
- Blow down samples under nitrogen

Honeybees 9/29/16

- Finished blowing down all samples under nitrogen gas so
  ~3mL were left behind.

- Filtered half of the honeybee samples following procedu
  from 7/28/16
  - These samples were placed back on the nitrogen evapora
    until dry, reconstituted with 1mL 10% MeOH:H_2O

Honeybees 9/30/16

- Filtered the other half of the honeybee samples following
  procedure from 7/28/16

10/3/16

- Honeybees
- Blow down more filtered samples under nitrogen until dry
- Reconstituted with 10% MeOH:H_2O

- Honeybees 10/4/16
- Blow down more filtered samples under nitrogen until dry

To Page No. ____

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	10/4/16

From Page No. ____

Honeybees 10/5/16

- Blow down more samples under nitrogen with a water bath at ~48° until dry

10/6/16

- Found 2 more honeybee samples that needed to be filtered following procedure on 7/28/16.

- Blow down more samples under nitrogen ~~with~~ until dry

- Added ~4 mL of MQ to several beebread samples
  ↳ vortexed until well-mixed
  ↳ place in freezer
  (Goal is freeze samples + place on freezer dryer to remove

Metalign for metabolomic / biomarker samples

- Used metalign to baseline correct, scale + align all chromatograms

With samples analyzed on the autospec
→ LF B/F 2014 (34) + BA B/F 2014 (n=34)
Fill in #1A - final + original folders
Fill in #2B - with your data as .cdf!
#4 → 1 #5 → 128,000 #6 → 1,000,000,000 #7 → 0.5
#8A → 1 #8B → 15 9 → ~~X~~5 → Hit #11 Run baseline

#12 → Autoscaling on total signal #13 → 0, 100 / 128000, 200 / 0, 0 / 0,
#14 → Pre-align Processing #15 → 20 #16 → 5, 3 #17 → 10, 5
#18 → ~~0~~ 19 → ~~0~~ → Hit: 20 (run scaling + Alignment)
→ Hit: 21 (multivariate → make-view)

To Page

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	10/6/16

Project No. _____

TITLE Metalign + beebread extraction

Book No. 4

From Page No. ____

10/6/16

- With sample analyzed on the TOF:
  - MB 10 (single 10's mix 10) n= 70
  - MB 10 - single n= 27
  - MB 10 - mixture n= 41
  - MB MAX (single max vs. mix max) n= 76
  - MB All - mixture (10 vs. MAX) n= 83
  - MB All - single (10 vs. max) n= 63

Fill in # 1A - final + original folders #2B - your folder w/ .cd
#4 → 1 #5 → 128,000 #6 → 1,000,000,000 #7 → 0.5 #8A -
#8B → 15 #9 → 75 Hit #11 → Run baseline

#12 - Autoscaling on total signal #13 0,100 / 128,000, 200 / 0,0 / 0,
#14 → Pre-align Processing #15 → 35 #16 → 5, 3 #17 → 10, 5
#18 → Ø #19 → Ø Hit 20: run scaling + alignment
Hit 21: Multivariate → make/view

Beebread 10/11/16

- Added 1 mL of MQ to beebread samples and vortexed until mixed
- Placed samples in the freezer overnight

10/12/16

Beebread

- Samples were removed from the freezer
- Removed caps + placed a kimwipe w/ rubberband on them
- Placed on freeze-dryer overnight to remove water

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	10/12/16

Project No. _____
Book No. 4 TITLE Beebread extraction method

rom Page No. ____

10/13/16

Beebread

- Removed beebread samples from the freeze-dryer and recapped samples.

Beebread SOP 10/18/16

1 - To each beebread sample add 6mL of EtOAc and vortex 10-15 min. until well mixed or freely moving

2 - Transfer supernatant to a pre-labeled glass cuvette

3 - To the original sample add 6mL of EtOAc + vortex for 1-2min + place on wrist-action shaker for 1 hr.

4 - Transfer supernatant to the ~~same~~ corresponding glass cuvette

5 - Repeat steps 3 + 4 once more for a total of 2 times with EtOAc

6 - Repeat steps 3+4 with MeOH to ensure all of pesticides have been removed

7 - Blow down sample until ~3mL ~~dry~~ under nitrogen gas with water bath 35° - 40°C.

~~8 - Once dry reconstitute with 1mL 10% MeOH:H_2O + vortex until well mixed.~~

~~8 - Filter samples~~

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	10/18/16

TITLE Beebread extraction

Project No. _____

Book No. 4

From Page No. ____

10/19/16

- Beebread
  - 4 beebread samples were extracted through steps 1-4 following procedure on pg. 54

10/20/16

Beebread
- Several more samples were extracted utilizing procedure on pg. 54
- slow going process

10/21/16

Beebread
- Several more samples were extracted following the pr on pg. 54 to part 5

10/24/16

Beebread
- several more samples were extracted following the proce on pg. 54 to part 5

10/25/16

Beebread
- several more samples were extracted following the procedure on pg. 54 to part 5

10/27/16

Beebread
- several more samples were extracted following the procedure on pg. 54. to part 5

To Page No

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	10/27/16

Project No. _____
Book No. 4 TITLE Neonic method + standards

From Page No. ____

10/28/16

- Using neonic standards from the EPA repository created a ~~new~~ ~1000 ppm mix standard in dichloromethane

Neonic standard		Purity
Thiamethoxam	- 9.27mg	98.9%
Clothianidin	- 8.61mg	99.6%
Nitenpyram	- 8.75mg	99.9%
Dinotefuran	- 8.80mg	99.9%
Acetamiprid	- 8.83mg	99.7%
Imidacloprid	- 8.34mg / 2 = 4.17	98.8
Thiacloprid	- 10.64mg / 5.32	99.6

put in 4 mL DCM (DG)

Transfered to bigger scintillation vial with 2 rinses of 2mL DCM

total 8mL at ~1000ppm

To Page No.

Witnessed & Understood by me, | Date | Invented by: | Date
Recorded by:

Project No. _____

TITLE Neonic GC method, beebread extraction

Book No. 4

From Page No. ____

10/31/16

Neonic standards/ GC

- Tried to develop a GC method for neonics

- Made up a 10ppm solution using 10uL of the stock solution into 990uL DCM + injected on GC

Neonic1 method

Ramp	Temp	Hold
	80	2 min
10	320	4 min

30 min

Potential peaks were identified

- had to create individual samples to verify Retention time Spectra

- ~~Took 100uL~~ ~~Standard stock from~~ ~~of~~ new neonic Mixture in ~~fridge~~
- Took old standard stock at 1000ppm in Methanol from freezer
  - Spiked 5uL into 2 sample vials for each standard
    - Both sets of vials were blown down under nitrogen until d
  - One set was reconstituted in EtOAc - 1mL
  - Set two was derivatized with 100uL of MSTFA + in the o at 60° for 1 hr.
  - All samples were placed on GC

Beebread samples

- All beebread samples were extracted following steps on pg. 54.

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	10/31/16

11/1/16

Neonic GC method

- Analyzed all individual standards that ran

Acetamiprid - 18.754 min MS (152, 126, 67, 166, 207, 221)

Thiacloprid - 22.131 min (207, 101, 126, 251)

Imidacloprid - 16.149 min (207, 126, 99)

Thiamethoxam - 15.200 min (132, 182, 212, 247)

Nitenpyram - 13.203 (67, 126, 169, 207)

Clothianidin ~ 9 min ? ?

Dinotefuran ?

- All of the samples that were derivatized gave similar values so ~~Rerun~~ the samples were probably not derivatized

Reran the EtOAc set one standards at 4ul to see if the peaks grow

Found - nitenpyram - 8.572 (?) (126, 155, 207)

clothianidin - 9.414 (?) (132, 207)

Dinotefuran - 17.295 (?) (207, 69, 83)

Beebread

- Finished blowing down samples until ~3mL were left in them under nitrogen gas

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/1/16

TITLE Neonic method GC + LC

Project No.

Book No. 4

From Page No.

11/2/16

Neonics GC

- Created a SIM method on GC

segment 1 - nitenpyram + clothianidin
MS - 126, 155, 207, 132

segment 2: 12 min nitenpyram
MS - 67, 126, 155, 207

segment 3: 14.5 min thiamethoxam, imidacloprid
MS - 132, 182, 97, 211
(also can use 212, 247, 126, 207)

segment 4: 17 min dinotefuran, nitenpyram, acetamiprid
MS - 207, 126, 152, 69, 83

segment 5: 21 min thiacloprid
MS - 101, 126

- GC single quad broke before I could run the SIM meth

LC/MS/MS - neonics

Matthew fixed the thermo!!

- Created a 10ppm solution by taking 10uL of ~1000ppm in DCM + blowing down under nitrogen until dry
- Reconstituted in 1mL 10% MeOH: H_2O
- Infused this on LC/MS/MS to obtain tube lens + collision en for each standard

Tune 161102_40_15_200_270

~~Acetamiprid~~ Dinotefuran	203 → 114 (-24V),	129 (-13V)	TL = 129
Acetamiprid	223 → 99 (-27V),	126 (-13)	TL = 163
Clothianidin	250 → 132 (-28V),	169 (-10V)	TL = 74
Thiacloprid	253 → 90 (-39V),	126 (-16V)	TL = 68
Imidacloprid	256 → 175 (-22V),	209 (-9V)	TL = 88
Thiamethoxam	292 → 181 (-15V),	211 (-10V)	TL = 53
Thiamethoxam D3	295 → 184 (-18V),	214 (-19V)	TL = 184
Nitenpyram	271 → 196 (-19V)	225 (-8V)	TL = 188

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Witnessed & Understood by me, | Date | Invented by: | Date 11/2/16

Recorded by:

Project No.
Book No. 4 TITLE Neonic standards LC

From Page No.

11/2/16

Made up a new set of standards for LC/MS/MS - Neonics

- 10 uL of standard mix in DCM of neonics made 10/28/16.

- Blow down dry under nitrogen gas & reconstituted in 1mL 10% MeOH : H_2O

- Made up a serial dilution

10ppm (z) = 1ppm (2mL)

z = 200uL in 1800 u

1ppm (x) = 0.5ppm (2mL)
0.5ppm (x) = 0.25ppm (2mL)
0.25ppm (x) = 0.125 (2mL)
0.125ppm (x) = 0.0625 (2mL)
0.0625ppm (x) = 0.03125 (2mL)
0.03125 ppm (x) = 0.015625 (2mL)
0.015625 ppm (x) = 0.00781 ppm (2mL)
0.00781 ppm (x) = 0.0039 ppm (2mL)
0.0039 ppm (x) = 0.00195 ppm (2mL)
0.00195 ppm (x) = 0.000976 ppm (2mL)
0.000976 ppm (x) - had 2mL total

X = 1mL

- Made up in 10% MeOH:H

- Spiked 2uL of thiamethoxam at 1000ppm into each stand

* 0.000976 ppm - spiked with 4uL of thiamethoxam

Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/2/16

TITLE Analyzed honeybees on LC/MS/MS

Project No.

Book No. 4

From Page No

11/2 - 11/4/14

Filename	Sample ID	Exp Method	Proc Method	Vial	Vol	Instrument
161102010	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102011	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102012	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102013	0.00097	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:2	20	Thermo
161102014	0.0019	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:3	20	Thermo
161102015	0.0039	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:4	20	Thermo
161102016	0.0078	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:5	20	Thermo
161102017	0.0156	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:6	20	Thermo
161102018	0.03125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:7	20	Thermo
161102019	0.0625	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:8	20	Thermo
161102020	0.125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:9	20	Thermo
161102021	0.25	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:10	20	Thermo
161102022	0.5	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:11	20	Thermo
161102023	1.0	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:12	20	Thermo
161102024	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102025	0.00097	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:2	20	Thermo
161102026	0.0019	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:3	20	Thermo
161102027	0.0039	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:4	20	Thermo
161102028	0.0078	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:5	20	Thermo
161102029	0.0156	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:6	20	Thermo
161102030	0.03125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:7	20	Thermo
161102031	0.0625	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:8	20	Thermo
161102032	0.125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:9	20	Thermo
161102033	0.25	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:10	20	Thermo
161102034	0.5	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:11	20	Thermo
161102035	1.0	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:12	20	Thermo
161102036	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102037	home apiary A1	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:13	20	Thermo
161102038	Brushy quarry A	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:14	20	Thermo
161102039	Doug McCracken B	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:15	20	Thermo
161102040	Pickle Station C	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:16	20	Thermo
161102041	Home yard 2A	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:17	20	Thermo
161102042	Home apiary 1B	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:18	20	Thermo
161102043	Home Yard 2b	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:19	20	Thermo
161102044	Doug McCracken A	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:20	20	Thermo
161102045	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102046	0.00097	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:2	20	Thermo
161102047	0.0019	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:3	20	Thermo
161102048	0.0039	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:4	20	Thermo
161102049	0.0078	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:5	20	Thermo
161102050	0.0156	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:6	20	Thermo
161102051	0.03125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:7	20	Thermo
161102052	0.0625	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:8	20	Thermo
161102053	0.125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:9	20	Thermo
161102054	0.25	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:10	20	Thermo
161102055	0.5	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:11	20	Thermo

Witness

Page No.

Recorded by:

Project No. _____
Book No. 4 TITLE Analyzed honeybees on LC/MS/MS

From Page No. ____ 11/2/16 - 11/4/16

Filename	Sample ID	Exp Method	Proc Method	Vial	Vol	Instrument
161102056	1.0	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:12	20	Thermo
161102057	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo
161102058	0.00097	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:2	20	Thermo
161102059	0.0019	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:3	20	Thermo
161102060	0.0039	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:4	20	Thermo
161102061	0.0078	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:5	20	Thermo
161102062	0.0156	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:6	20	Thermo
161102063	0.03125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:7	20	Thermo
161102064	0.0625	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:8	20	Thermo
161102065	0.125	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:9	20	Thermo
161102066	0.25	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:10	20	Thermo
161102067	0.5	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:11	20	Thermo
161102068	1.0	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:12	20	Thermo
161102069	blank	C:\Xcalibur\methods\Neonics_2016	C:\Xcalibur\methods\Neonics	A:1	20	Thermo

DPAG

Using the corrected values for each standard in the standard mix that was created on 10/28/16, ~~able~~ able to quantify the 7 neonics in the 8 bee incident 2016 samples.

- The first set of standards that ran were off along with several from the second set. The 3rd + 4th set were spot on for each standard + mainly used for linear regression

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	

TITLE Beebread extraction

Project No.

Book No. 4

From Page No. ____

Beebread 11/7/16

Filtering beebread samples after they were extracted + blown down

- Changed all of the liners + rinsed with MeOH on vacuum man
  - Replace the lid on top after adding all new test tubes labeled
- Put new syringe + filter on top of manifold
- Vortex beebread sample until well mixed for at least 30 se
- Add to syringe filter with corresponding name + # using a gle pipette
- Add a small amount of MeOH to each sample + vortex for at least 10 sec → add to syringe filter
- Add a small amount of MeOH to each sample + vortex for at least 10 sec → add to syringe filter
- Add 2/3 MeOH + 1/3 ratio EtOAc to each sample + vortex for at least 10 sec → add to syringe filter
- Add 2/3 MeOH + 1/3 ratio EtOAc to each sample + vortex for at least 10 sec → add to syringe filter
- Use syringe plunger only to push down solvent
- when done place test tube under steady stream of nitrog + evaporate to dryness

11/8/16

Beebread

- Following the filtering procedure from 11/7/16 the remaining beebread samples were filtered.
- Placed under nitrogen with a water bath @ 35°C.

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/8/16

Project No.
Book No. 4
TITLE Honey extraction methods

From Page No.

11/8/16

Honey

- Trying 2 different extraction methods for honey
  - one is by Sánchez - Hernandez et al 2016)

1 - In triplicates, weighed out 5grams of honey into a new 15mL tube.
2 - Spiked with 2uL of 1000ppm thiamethoxam
3 - Spiked with 20uL of 10ppm nemic std in MeOH)
4 - Added 10mL of MQ water and vortex until well mixed
  - On vacuum manifold replace inlet liners & cleaned w/ Me
5 - Place a new cartriage (strata-X 33um 200mg/6cc) prelabe on top of vacuum manifold
6 - Precondition the cartridge with 6mL MeOH, 6mL MQ
7 - Load sample on to cartridge
8 - After sample passes through place prelabeled test tubes inside manifold
9 - Wash cartridge + centrifuge tube with 2mL of 70% MeOH:H_2O
10 - Elute with 2mL 45% ACN:H_2O into prelabeled test tu
11 - Place sample on speedvac overnight.

Method #2 (Pirard, C. Agilent Technologies)

1 - Weighed out 1gram of honey in triplicates
2 - Spiked with 2uL of thiamethoxam at 1000ppm
3 - Spiked with 20uL of nemic standard at 10ppm MeOH
4 - Added 1.25mL of H_2O + 2.5mL of acetone
5 - Added 1.25mL of NaCl solution (2g/10mL)
6 - Vortex until well mixed
7 - Apply sample to a preconditioned SPE (strata-X, 33um, 200mg/
8 - Allow for 15min to pass to allow absorption to take place
9 - Elute sample with 18mL of EtOAc into pre-labeled test tube
10 - Evaporate to dryness under nitrogen
11 - Reconstitute in 10% MeOH at 1mL

- only got to Step 6

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/8/16

TITLE Honey extraction methods

Project No.

Book No. 4

From Page No.

11/9/16

Honey

Method 1

- Removed from speedvac
- Added 1mL of 10% MeOH:H_2O + vortex until dissolved (had to let sit for (1hr). Transfered to a 2mL centrifuge tube w/ filter @ 14000rpm for 15min. Transfered to pre-labeled GCV

Method 2

- Finished steps 7, 8, + 9

Beebread

- Blow down all the samples under nitrogen gas until dry
- Reconstitute with 1mL of 10% MeOH:H_2O + let sit for 1hr + vortex until well mixed

11/10/16

Method 2 for honey - Finished step 10 ~~for~~ blowing down under nitrogen

Honey Method 3

- Repeated the above procedure for honey #1 except the 2mL 70% MeOH:H_2O + 2mL 45% ACN:H_2O were captured in 2 different test tubes to determine if any neonics were present in the 70% MeOH fraction
- Got through eluting with different solvents and placed on speedvac on Monday

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		Recorded by:	11/10/16

Project No.
Book No. 4 TITLE Honey extraction

From Page No.

11/14/16

Honey Method 2
- Reconstituted samples in 200uL of 10% ACN:H_2O + vortexed until well mixed
- Transferred samples to a pre-labeled GC vial with an insert

Honey Method 3 - separate
- Placed samples on speedvac for several hours to dry off solvent
- Samples were reconstituted in 1mL of 10% ACN:H_2O
- Vortex until well mixed
- Transferred to a 2mL centrifuge tube with a 0.2um filter for 15min @ 14000rpm
- Solvent was transferred to a pre-labeled GC vial

70% MeOH labeled 1-2, 2-2, 3-2
45% ACN labeled 1-3, 2-3, 3-3

Procedural blank was done in triplicates following the first honey method to see if there is any contaminants in the honey and how ~~well~~ clean the environment is.
Honey Method #1 is on pgs 64 + 65

- Placed samples on speedvac overnight

- Ran samples on Thermo pg. 72 - dead bee trap
11/15/16

- Removed samples from speedvac. procedural blanks for honey

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/15/16

TITLE Honey Extraction

Project No.

Book No. 4

From Page No.

11/16/16

Honey

- Weighed out 5 grams of honey into 15mL centrifuge tub for 2 sets of standards (11 standards)

- Made up new working solution of 10 ppm neonic standard Took 10uL of 1000ppm neonic std from 10/28/16 into a 2 vial + blow down under nitrogen + reconstitute in 10% MeOH:H_2O

- All honey are spiked with the following standard

10 ppm (A) = 1mL (1ppm) A = 100uL

10 ppm (B) = 1mL (0.5ppm) B = 50uL

10ppm (C) = 1mL (0.25ppm) C = 25uL

10ppm (D) = 1mL (0.125 ppm) D = 12.5uL

10ppm (E) = 1mL (0.0625ppm) E = 6.25uL

10ppm (F) = 1mL (0.03125ppm) F = 3.125uL

0.15625ppm (G) = 1mL (0.015625ppm) G = 100uL

0.15625ppm (H) = 1mL (0.0078125ppm) H = 50uL

0.15625ppm (I) = 1mL (0.003906ppm) I = 25uL

0.15625ppm (J) = 1mL (0.00195 ppm) J = 12.5uL

0.15625ppm (K) = 1mL (0.000976ppm) K = 6.25

10ppm (X) = 0.15625ppm (2mL) X = 0.03125mL

31.25uL

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/16/16

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11/16/16

Honey Method #1

- Following the spike of neonic standard each standard was also spiked with 2uL of 1000ppm thiamethoxam D3
- Each standard had 10mL MQ added to each sample
- Vortex until well mixed

11/17/16

- Honey Method #1
  - SPE cartridges were pre-conditioned with 5mL MeOH + 5mL MQ
  - Honey sample was passed through the cartridge
  - Test tubes were placed under each sample to catch the 2mL wash - 70% MeOH:H_2O (placed in test tube to clean sample
  - Cartridges were eluted with 4mL 45% ACN:H_2O
  - Samples were placed on speedvac overnight.

11/18/16

Honey Method #1

- The second set of standards were extracted following the method from 11/17/16
- Samples were removed from speedvac and reconstituted with 1mL 10% ACN:H_2O
- Let set for 1 hr + vortex until well mixed
- Samples were placed in a prelabeled 2mL centrifuge tube w/ filter - centrifuged @ 15000 for 10min.
- Bottom sample was then transferred to a 2mL GC vial pack
- Samples were placed in freezer until analyzed on LC/MS

To Page

TITLE Malaoxon standards

Project No. _____

Book No. 4

From Page No. ____

11/21/16

- Honey samples/standards
- Removed honey stds from speedvac + reconstituted in 10% ACN:H_2O w/ 1
- Followed procedure from 11/18/16

Malaoxon

- Weighed out 4.56 mg of malaoxon standard from Sigma-Aldrich 98.1% purity w/ Batch # SZBF146XV
- Dissolved malaoxon in 4.56 mL of EtOAc to get 1000 ppm standard
- Took a 4 uL aliquot and put it into 2 mL to get a 2 ppm standard to run on GCMS
- Ions - 99, 109, 127

- Made a serial dilution for a calibration curve of malaoxon

1000ppm	(z)	=	2mL	(2ppm)
2ppm	(x)	=	2mL	(1ppm)
1ppm	(x)	=	2mL	(0.5 ppm)
0.5ppm	(x)	=	2mL	(0.25ppm)
0.25ppm	(x)	=	2mL	(0.125ppm)
0.125ppm	(x)	=	2mL	(0.0625)
0.0625	(x)	=	2mL	(0.03125)
0.03125	(x)	=	2mL	(0.015625)
0.015625	(x)	=	2mL	(0.0078)
0.0078	(x)	=	2mL	(0.0039)

z = 4 uL

x = 1 mL

solvent was EtOAc

- All standards were spiked with 3 1/3 uL of 1000ppm tetraconazole

- All of Robin's (RJV) summer 2015 frogs were reconstituted in 1mL EtOAc to be analyzed on GC/MS

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	11/21/16

Project No.
Book No. 2
TITLE Inhive nursebees Thermo
11/14/16 - 11/21/16

From Page No.

File name	Sample	Path	Method	Data Processing	Vol	Instrument
161114001	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114002	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114003	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114004	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114005	0.00097	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114006	0.00190	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114007	0.00390	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114008	0.00780	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114009	0.01563	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114010	0.03125	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114011	0.06250	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114012	0.12500	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114013	0.25000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114014	0.50000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114015	1.00000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114016	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114017	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114018	0.00097	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114019	0.00190	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114020	0.00390	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114021	0.00780	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114022	0.01563	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114023	0.03125	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114024	0.06250	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114025	0.12500	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114026	0.25000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114027	0.50000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114028	1.00000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114029	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114030	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114031	Home apiary 1a_1	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114032	brushy quarry a	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114033	Doug McCracken B_2	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114034	Pickle station	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114035	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114036	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114037	Home yard 2b_1	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114038	Home apiary 1A_2	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114039	Home yard 2b_2	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114040	Doug McCracken B_1	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114041	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114042	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114043	IH NB 85	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114044	IH NB 91	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114045	IH NB 103	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114046	IH NB 90	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114047	IH NB 99	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114048	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114049	BLANK	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114050	IH NB 93	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114051	IH NB 107	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS

Witnessed & Understood by me, | Date | Invented by: | Date
Recorded by:

Project No.

TITLE IH nurscbees Thermo

Book No. 4

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11/14/16 - 11/21/16

File name	Sample	Path	Method	Data Processing	Vol	Instrument
161114052	IH NB 97	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114053	IH NB 104	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114054	IH NB 86	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114055	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114056	BLANK	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114057	IH NB 112	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114058	IH NB 106	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114059	IH NB 88	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114060	IH NB 109	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114061	IH NB 95	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114062	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114063	BLANK	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114064	IH NB 101	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114065	IH NB 105	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114066	IH NB 98	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114067	IH NB 110	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114068	IH NB 92	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114069	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114070	BLANK	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114071	IH NB 100	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114072	IH NB 94	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114073	IH NB 96	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114074	IH NB 87	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114075	IH NB 108	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114076	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114077	BLANK	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114078	IH NB 89	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114079	IH NB 111	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114080	IH NB 102	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114081	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114082	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
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161114084	0.00190	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114085	0.00390	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114086	0.00780	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114087	0.01563	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114088	0.03125	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114089	0.06250	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114090	0.12500	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114091	0.25000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114092	0.50000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114093	1.00000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114094	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114095	QAQC	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114096	0.00097	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114097	0.00190	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114098	0.00390	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114099	0.00780	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114100	0.01563	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114101	0.03125	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161114102	0.06250	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)

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Witnessed & Understood by me, | Date | Invented by: | Date 12/2/16

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TITLE 1H Nurseberes Thermo
11/14/16 - 11/21/16

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File name	Sample	Path	Method	Data Processing	Vol	Instrument
161114103	0.12500	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114104	0.25000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114105	0.50000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114106	1.00000	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/MS
161114107	blank	Xcalibur/Data/Neonics/161114	Neonic_2016	Neonic	20	Thermo (LC/MS/M

NEONIC METHOD FOR THERMO

TSQ Quantum Instrument Method
Creator: Quantum Last modified: 11/14/2016 10:04:29 AM by Quantum
TSQ MS Method Settings:
Method Type: Regular Method
MS Run Time (min): 30.00
Segment 1
Duration (min) 30.00
Scan Events 1
Segment 1:
Tune Method C:\Xcalibur\methods\Tune 161114_40_15_200_270_neonic.TSQTune
Chrom filter: 5.0
Q2 Gas Pressure: 1.5
Syringe Pump: Off
Scan Events:
1: + c SRM Skimmer Offset 10, Micro Scans 1, Accurate Mass Off,

Parent	Center	Width	Time	CE	Q1 PW	Q3 PW	Tube Lens
203.000	114.000	0.500	0.300	11	1.00	1.00	Tuned Value
203.000	129.000	0.500	0.300	10	1.00	1.00	Tuned Value
223.000	99.000	0.500	0.300	33	1.00	1.00	Tuned Value
223.000	126.000	0.500	0.300	20	1.00	1.00	Tuned Value
250.000	132.000	0.500	0.300	10	1.00	1.00	Tuned Value
250.000	169.000	0.500	0.300	9	1.00	1.00	Tuned Value
253.000	90.000	0.500	0.300	34	1.00	1.00	Tuned Value
253.000	126.000	0.500	0.300	19	1.00	1.00	Tuned Value
256.000	175.000	0.500	0.300	17	1.00	1.00	Tuned Value
256.000	209.000	0.500	0.300	17	1.00	1.00	Tuned Value
271.000	196.000	0.500	0.300	18	1.00	1.00	Tuned Value
271.000	225.000	0.500	0.300	10	1.00	1.00	Tuned Value
292.000	181.000	0.500	0.300	18	1.00	1.00	Tuned Value
292.000	211.000	0.500	0.300	9	1.00	1.00	Tuned Value
295.000	184.000	0.500	0.300	21	1.00	1.00	Tuned Value
295.000	214.000	0.500	0.300	17	1.00	1.00	Tuned Value

Syringe pump not in use
Divert Valve: not used during run
Accurate Mass Settings
No lock masses entered

Time	%B	%D	Flow Rat
0	98	2	2
0.5	98	2	2
10	65	35	2
14	2	98	2
17	2	98	2
17.1	98	2	2
30	98	2	2

B = H_2O w/ 0.05% FA + Ammonium Acet

D = MeOH w/ 0.05% FA Ammonium Acet

DAB

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- Ran the dead bee trap (honey/nurse bees) on the Thermo LC/MS/MS
- Needle broke so it had to be replaced 2/3 way through
- Pressure dropped after ~~several~~ 40 samples had to rerun several
- Arm malfunctioned halfway through second set of samples

- Both ions for each neonic was quantified for each standard + sample

- 2 sets of standards
- Samples
- Halfway through 1 set of standards
- Samples
- 2 sets of standards
- QAQC + blank every 5th sample

- Standards & QAQC were made up fresh following the method on pg 60

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File name	Sample	Path	Method	Data Processing	Vol	Instrument
161209001	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209002	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209003	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209004	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209005	0.00097	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209006	0.0019	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209007	0.0039	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209008	0.0078	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209009	0.015625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209010	0.03125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209011	0.0625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209012	0.125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209013	0.25	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209014	0.5	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209015	1	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209016	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209017	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209018	0.00097	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209019	0.0019	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209020	0.0039	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209021	0.0078	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209022	0.015625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209023	0.03125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209024	0.0625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209025	0.125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209026	0.25	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209027	0.5	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209028	1	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209029	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209030	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209031	BLANK	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209032	IH NB 88	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209033	IH NB 109	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209034	IH NB 95	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209035	DBT 26	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209036	DBT 64	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209037	DBT 49	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209038	DBT 78	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209039	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209040	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161209041	DBT 13	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211001	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211002	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211003	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211004	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211005	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211006	DBT 36	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211007	DBT 43	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211008	DBT 84	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211009	DBT 20	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211010	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211011	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211012	DBT 76	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211013	DBT 92	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)

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File name	Sample	Xcalibur/Data/Neonics/1601209	Method	Data Processing	Inj	Instrument
161211014	DBT 22	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211015	DBT 5	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211016	DBT 74	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211017	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211018	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211019	DBT 18	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211020	DBT 45	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211021	DBT 57	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211022	DBT 97	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211023	DBT 28	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211024	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211025	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211026	DBT 62	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211027	DBT 24	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211028	DBT 53	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211029	DBT 69	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211030	DBT 89	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211031	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211032	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211033	DBT 12	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211034	DBT 66	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211035	DBT 86	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211036	DBT 38	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211037	DBT 63	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211038	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211039	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211040	DBT 72	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211041	DBT 16	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211042	DBT 50	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211043	DBT 27	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211044	DBT 80	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211045	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211046	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211047	DBT 21	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211048	DBT 94	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211049	DBT 40	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211050	DBT 71	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211051	DBT 32	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211052	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211053	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211054	DBT 33	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211055	DBT 52	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211056	DBT 88	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211057	DBT 44	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211058	DBT 19	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211059	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211060	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211061	DBT 9 OR 6	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211062	DBT 75	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211063	DBT 56	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211064	DBT 35	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211065	DBT 8	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211066	DBT 3	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211067	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)

Witnessed

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File name	Sample		Method	Data Processing	Inj	Instrument
161211068	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211069	0.00097	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211070	0.0019	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211071	0.0039	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211072	0.0078	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211073	0.015625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211074	0.03125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211075	0.0625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211076	0.125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211077	0.25	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211078	0.5	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211079	1	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211080	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211081	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211082	DBT 68	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211083	DBT 100	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211084	DBT 51	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211085	DBT 83	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211086	DBT 17	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211087	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211088	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211089	DBT 59	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211090	DBT 77	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211091	DBT 6 OR 9	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211092	DBT 42	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211093	DBT 70	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211094	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211095	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211096	DBT 98	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211097	DBT 54	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211098	DBT 31	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211099	DBT 47	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211100	DBT 95	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211101	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211102	BLANK	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211103	DBT 48	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211104	DBT 2	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211105	DBT 15	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211106	DBT 29	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211107	DBT 82	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211108	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211109	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211110	DBT 67	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211111	DBT 79	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211112	DBT 23	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211113	DBT 37	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211114	DBT 14	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211115	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211116	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211117	DBT 73	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211118	DBT 81	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211119	DBT 39	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211120	DBT 65	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211121	DBT 11	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)

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File name	Sample	Xcalibur/Data/Neonics/1601209	Method	Data Processing	Inj	Instrument
161211122	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211123	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211124	DBT 87	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211125	DBT 55	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211126	DBT 99	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211127	DBT 4	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211128	DBT 58	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211129	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211130	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211131	DBT 30	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211132	DBT 34	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211133	DBT 85	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211134	DBT 61	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211135	DBT 96	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211136	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211137	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211138	DBT 25	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211139	DBT 1	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211140	DBT 91	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211141	DBT 60	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211142	DBT 46	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211143	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211144	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211145	DBT 93	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211146	DBT 10	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211147	DBT 90	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211148	DBT 41	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161211149	DBT 7	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215001	BLANK	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215002	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215004	0.00097	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215005	0.0019	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215006	0.0039	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215007	0.0078	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215008	0.015625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215009	0.03125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215010	0.0625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215011	0.125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215012	0.25	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215013	0.5	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215014	1	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215016	QAQC	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215017	0.00097	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215018	0.0019	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215019	0.0039	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215020	0.0078	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215021	0.015625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215022	0.03125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215023	0.0625	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215024	0.125	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215025	0.25	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215026	0.5	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215027	1	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)
161215028	blank	Xcalibur/Data/Neonics/1601209	Neonic_2016	Neonic	20	Thermo (LC/MS/MS)

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Vial	Filename	Sample ID	Exp Method	Proc Method	Inj	Instrument
A:1	170103001	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
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A:2	170103004	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:3	170103005	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:4	170103006	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:5	170103007	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:6	170103008	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:7	170103009	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:8	170103010	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:9	170103011	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:10	170103012	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:11	170103013	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:12	170103014	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:13	170103015	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:1	170103016	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:2	170103017	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:3	170103018	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:4	170103019	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:5	170103020	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:6	170103021	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:7	170103022	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:8	170103023	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:9	170103024	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:10	170103025	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:11	170103026	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:12	170103027	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
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A:15	170103032	NB 3-83	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:16	170103033	NB 5-104	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:17	170103034	NB 5-110	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:18	170103035	NB 3-76	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
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a:1	170103037	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:19	170103038	NB 5-116	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:20	170103039	NB 2-47	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:21	170103040	NB 5-123	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:22	170103041	NB 4-91	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:23	170103042	NB 3-86	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
a:2	170103043	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
a:1	170103044	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:24	170103045	NB 3-64*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:25	170103046	NB 5-121	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

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Vial	Filename	Sample ID	Exp Method	Proc Method	Inj	Instrument
A:26	170103047	NB 3-68	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:27	170103048	NB 3-72	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:28	170103049	NB 2-52	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
a:2	170103050	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
a:1	170103051	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:29	170103052	NB 3-89	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:30	170103053	NB 4-97	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:31	170103054	NB 5-117	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:32	170103055	NB 5-125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:33	170103056	NB 4-101	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
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A:34	170103059	NB 2-49	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:35	170103060	NB 2-39	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:36	170103061	NB 4-93	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:37	170103062	NB 5-113	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:38	170103063	NB 4-92	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
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a:1	170103065	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:39	170103066	NB 3-75	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:40	170103067	NB 3-70	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:1	170103068	NB 5-129	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:2	170103069	NB 3-65	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:3	170103070	NB 2-56	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
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B:4	170103073	NB 3-67	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:5	170103074	NB 5-108	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:6	170103075	NB 2-63	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:7	170103076	NB 2-42	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:8	170103077	NB 3-87	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
a:2	170103078	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
a:1	170103079	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:9	170103080	NB 4-96	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:10	170103081	NB 5-112*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:11	170103082	NB 3-82	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:12	170103083	NB 4-90	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:13	170103084	NB 5-128	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
B:14	170103085	NB 2-54	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:1	170103086	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:2	170103087	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:3	170103088	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:4	170103089	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:5	170103090	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:6	170103091	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:7	170103092	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

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Vial	Filename	Sample ID	Exp Method	Proc Method	Inj	Instrument
A:8	170103093	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:9	170103094	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:10	170103095	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:11	170103096	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:12	170103097	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:13	170103098	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:1	170103099	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:2	170103100	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:3	170103101	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:4	170103102	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:5	170103103	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:6	170103104	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:7	170103105	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:8	170103106	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:9	170103107	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:10	170103108	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:11	170103109	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:12	170103110	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:13	170103111	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
A:13	170103112	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

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Filename	Vial	Sample ID	Exp Method	Proc Method	Inj	Instrument
170106001	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106002	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106003	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106004	A:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106005	A:3	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106006	A:4	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106007	A:5	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106008	A:6	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106009	A:7	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106010	A:8	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106011	A:9	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106012	A:10	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106013	A:11	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106014	A:12	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106015	A:13	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106016	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106017	A:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106018	A:3	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106019	A:4	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106020	A:5	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106021	A:6	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106022	A:7	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106023	A:8	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106024	A:9	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106025	A:10	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106026	A:11	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106027	A:12	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106028	A:13	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106029	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106030	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106031	a:14	NB 2-51	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106032	A:15	NB 4-102	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106033	A:16	NB 5-115	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106034	A:17	NB 5-106	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106035	A:18	NB 2-50	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106036	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106037	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106038	A:19	NB 3-85*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106039	A:20	NB 3-74	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106040	A:21	NB 5-127	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106041	A:22	NB 5-131	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106042	A:23	NB 5-110	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106043	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106044	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106045	A:24	NB 4-95	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106046	A:25	NB 3-80	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

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Filename	Vial	Sample ID	Exp Method	Proc Method	Inj	Instrument
170106047	A:26	NB 2-41	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106048	A:27	NB 5-109	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106049	A:28	NB 5-124	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106050	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106051	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106052	A:29	NB 4-100	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106053	A:30	NB 5-114	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106054	A:31	NB 3-66	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106055	A:32	NB 2-59	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106056	A:33	NB 5-119	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106057	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106058	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106059	A:34	NB 3-71	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106060	A:35	NB 2-46	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106061	A:36	NB 5-120	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106062	A:37	NB 5-122	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106063	A:38	NB 4-92	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106064	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106065	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106066	A:39	NB 2-62	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106067	A:40	NB 5-111	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106068	B:1	NB 5-105	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106069	B:2	NB 3-84	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106070	B:3	NB 3-76	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106071	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106072	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106073	B:4	NB 2-43	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106074	B:5	NB 3-69	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106075	B:6	NB 3-78	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106076	B:7	NB 3-73*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106077	B:8	NB 4-103	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106078	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106079	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106080	B:9	NB 2-60	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106081	B:10	NB 4-94	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106082	B:11	NB 2-40	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106083	B:12	NB 5-118	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106084	B:13	NB 5-130* SP	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106085	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106086	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106087	B:14	NB 3-77	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106088	B:15	NB 3-81	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106089	B:16	NB 4-99	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106090	B:17	NB 2-55*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106091	B:18	NB 2-53	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106092	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

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Filename	Vial	Sample ID	Exp Method	Proc Method	Inj	Instrument
170106093	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106094	B:19	NB 3-88	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106095	B:20	NB 2-57	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106096	B:21	NB 5-107	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106097	B:22	NB 4-98	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106098	B:23	NB 5-126	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106099	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106100	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106101	B:24	NB2-45	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106102	B:25	NB 3-79	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106103	B:26	NB 2-48*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106104	b:27	NB 5-106	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106105	B:28	NB 2-50	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106106	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106107	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106108	B:29	NB 3-85*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106109	B:30	NB 3-74	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106110	b:1	NB 4-92	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106111	B:2	NB 5-111	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106112	B:3	NB 2-55*2	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106113	a:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106114	a:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106115	B:4	NB 2-53	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106116	b:5	NB 3-88	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106117	B:6	NB 2-57	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106118	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106119	A:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106120	A:3	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106121	A:4	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106122	A:5	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106123	A:6	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106124	A:7	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106125	A:8	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106126	A:9	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106127	A:10	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106128	A:11	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106129	A:12	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106130	A:13	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106131	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106132	A:2	qaqc	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106133	A:3	0.00097	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106134	A:4	0.0019	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106135	A:5	0.0039	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106136	A:6	0.00781	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106137	A:7	0.015625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106138	A:8	0.03125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

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Recorded by:

1/9/17

No.

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TITLE Nucbox nursebees - Thurmo

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Filename	Vial	Sample ID	Exp Method	Proc Method	Inj	Instrument
170106139	A:9	0.0625	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106140	A:10	0.125	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106141	A:11	0.25	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106142	A:12	0.5	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106143	A:13	1.0	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106144	A:1	blank	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106145	B:1	NB 3-79	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS
170106146	B:2	NB 3-88	C:\Xcalibur\methods\Neonics_2016.meth	C:\Xcalibur\methods\Neonics	20	Thermo_LC/MS/MS

DAG

- Weighed out 13 - 5 grams of honey into 15 mL test tubes to be spiked with neonic standards for a calibration curve to be ran with honey + beebread samples

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Witnessed & Understood by me,	Date	Invented by:	Date
		Recorded by:	1/9/17