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Governor's Office of Homeland Security & Emergency Preparedness

INCIDENT ACTION PLAN - 12

Incident Name: Tulane National Primate Research Center 15-005

Unified Command:

CDC - RADM Scott Deitchman

GOHSEP - Dir. Kevin Davis

LA DHH - Dr. Jimmy Guidry

St. Tammany - Dir. Dexter Accardo

Approved:

Operational Period:

Date: April 3, 2015

Time: 4/03/2015 - 0900hrs to 4/10/2015 - 0900hrs

St. Tammany Parish EOC
510 E. Boston Street
Covington, La. 70433
Office. 985-898-2359

St. Tammany Parish EOC - GPS Coordinates
30.476016, -90.095449

Prepared By	C.R.Simoneaux	Signature		Date		Time	

90040932

Governor's Office of Homeland Security & Emergency Preparedness

INCIDENT OBJECTIVES		Incident Number	15-005	ICS 202
Incident Name	Tulane National Primate Research Center	Operational Period	4/03/2015 0900hrs to 4/10/2015 0900hrs	

Objectives

	** Note - some field operations have resumed, including rodent, feral cat, wildlife and NHP sampling and testing. ** ** TNPRC, Local, State and Federal Agencies are continuing work on developing short and long term monitoring plans**
1	**Investigation process - On Campus: To identify, isolate, remediate and prevent further tranmission of Bp on campus grounds.**
	Sampling Objectives:
1.1	To conduct serological sampling to determine potential parameters of transmission exposure.
	Status: *Human :* *LA DHH Updates as of 4/2/15 (updates are underlined) -* *Batch 3 – On March 13,2015 4 blood samples were sent to CDC for testing. One was a re-test from the March 11, 2015 batch; this samples continues to show weak reactivity with no change in titer. All other samples were negative.* *Batch 4 – On March 20, 2015 30 blood samples were collected by OPH. Of the 30, 29 specimens were shipped to CDC to arrive on March 24, 2015. Of the 30, one specimen is to be banked at the OPH lab. OPH/Epi is waiting on the results of these specimens. On March 26, CDC provided the following results – of the 29 samples, 27 were negative and 2 were inconclusive and require a re-draw. OPH and Tulane have scheduled the 2 redraws to occur on April 7, 2015 .* *Batch 5 – On March 25, 2015 1 blood sample was collected by OPH. OPH is to confirm with individual whether sample will be tested by CDC or banked at OPH lab. CDC provided the result on March 31. Result was negative.* *Non-Human Primate (NHP):* *CDC Updates as of 3/31/15 -* *Latest Animal Results* *· 1/57 samples reactive JC21 (1:10240)* *· J21 was in the clinic on Dec 16th and then Jan 29th onward. From field cage J/C17* *· We shared results with Tulane* *Current status of animal testing* *· 3 culture-confirmed infections (symptomatic)* *· 3 asymptomatic seroconversions* *· 9 asymptomatic NHPs pending further investigation* *ANIMAL TESTING - IHA* *BIG PICTURE* *· There are 4107 animals in the facility. We have tested 681 of them (16.6%).* *· There have been 840 animals in the clinic between Oct 15 and Feb 15. We have tested 477(56.7%), 427 (89.5%) are from current samples (drawn after 2/2/15).*
1.2	To conduct air sampling to determine potential of aerosolized transmission.
	Status: Completed. All test results were negative for Bp. Additional short term and long term environmental monitoring plans are being developed.
1.3	To conduct soil sampling within the cages to validate spread of Bp in cage-soil.
	Status: All test results were negative for Bp. Additional short term and long term environmental monitoring plans are being developed.

Prepared by	C.R.Simoneaux	Signature		Date		Time	
Approved by		Signature		Date		Time	

Governor's Office of Homeland Security & Emergency Preparedness

INCIDENT OBJECTIVES		Incident Number	15-005	ICS 202 (page 2)
Incident Name	Tulane National Primate Research Center	Operational Period	4/03/2015 0900hrs to 4/10/2015 0900hrs	

Objectives (continued)

1.4	To conduct soil sampling outside the cages to determine if Bp has spread beyond enclosures
	Status: All test results were negative for Bp. Additional short term and long term environmental monitoring plans are being developed.
1.5	To conduct water sampling around the compound and at discharge points to determine if Bp is growing in water discharges.
	Status: All test results were negative for Bp. Additional short term and long term environmental monitoring plans are being developed.
	Remediation Objectives:
1.6	To conduct remediation of enclosures where infected animals were - or are - located.
	Status: Awaiting status update from Tulane.
1.7	To conduct remediation outside the cages based on soil sampling results.
	Status: Awaiting status update from STWG and Tulane based on additional short term and long term environmental monitoring plans being developed.
1.8	To identify potential mitigation activities should air and/or water samples come back positive.
	Status: Awaiting status update from STWG based on additional short term and long term environmental monitoring plans being developed.
2	**Surrounding Area - Off Campus: To conduct risk assessment(s) within a meaningful vicinity outside the compound to ascertain need for further sampling wildlife, livestock, and other potential at-risk animals.**
2.1	To conduct risk survey of animals in the area and determine whether these animals require further testing.
	Status: USDA/APHIS is developing a risk assessment and additional short term and long term monitoring plan. Some wildlife, rodent and feral cat sampling is underway with samples being sent to the USDA National Veterinary Services Laboratory (NVSL). Initial wildlife samples sent for testing were all negative for Bp. Additional samples have been sent. Awaiting test results for review by the Science Technical Working Group (STWG).
2.2	To utilize findings from the investigative process to determine need for further mitigation activities outside the compound.
	Status: USDA/APHIS is developing a risk assessment and additional short term and long term monitoring plan.
3	**Public Communications**
3.1	To formalize JIC members under CDC lead.
	Status: Completed but under GOHSEP lead.
3.2	To develop Frequently Asked Questions document in readiness for public dissemination.
	Status: A public meeting was held in St. Tammany Parish on 3/31/15. The latest media release is attached.

Prepared by	C.R.Simoneaux	Signature		Date		Time	
Approved by		Signature		Date		Time	

Governor's Office of Homeland Security & Emergency Preparedness

INCIDENT OBJECTIVES		Incident Number	15-005	ICS 202 (page 3)
Incident Name	Tulane National Primate Research Center	Operational Period	4/03/2015 0900hrs to 4/10/2015 0900hrs	

Objectives (continued)

4	**Science Technical Working Group - To advise the Unified Command on Interpretation of Sampling and Testing Results, and to Make Recommendations on Further Actions with Investigations, Remediation and Response (Objectives 1 and 2).**
4.1	Review, interpret and advise the UC on human blood test results and need for additional actions.
	Status: Initial test results are being reviewed. Awaiting additional test results for review.
4.2	Review, interpret and advise the UC on animal blood test results and need for additional actions.
	Status: Initial test results are being reviewed. Awaiting additional test results for review.
4.3	Review, interpret and advise the UC on air, water, and soil test results and need for additional actions.
	Status: Initial test results are being reviewed and additional short term and long term environmental monitoring plans are being developed.

Operational Period Command Emphasis
(Priorities, Key Decisions/Directions)

Expedient development and implementation of testing and response plans is the Unified Command emphasis, while at all times making decisions which are in the best interest of worker and public safety. Any issues which arise to impede this effort will be immediately communicated to the Unified Command for assistance and resolution. Full compliance with animal health and welfare regulations will be observed, and efforts will be made to minimze impacts to animal welfare.

NWS Weather Forecast for Covington, La. as of 4/02/2015 at 0900hrs CST

National Weather Service Office Slidell, La. 985-649-0429 http://www.srh.noaa.gov/lix/

Safety Message / Site Safety Plan

Safety of response personnel and the general public is paramount. Primary focus will be on strict adherence to safety rules and regulations and proper use of PPE. Any safety concerns, incidents or injuries must be immediately reported to a supervisor and the incident Safety Officer.

Is a Site Safety Plan Required?	X Yes	No	Location of Site Safety Plan: On file with Safety Officer and Planning Section

Incident Action Plan Components (check if attached)

x	Incident Action Plan Cover (ICS-200)		Medical Plan (ICS-206)
x	Incident Objectives (ICS-202)	x	Site Safety Plan (ICS-208)
x	Organization Assigment List (ICS-203)	x	Cleaning and Disinfecting Vehicles Plan
	Assignment List (ICS-204)	x	Incident Map(s)
x	Communications Plan (ICS-205T)		

Prepared by	C.R.Simoneaux	**Signature**		**Date**		**Time**	
Approved by		**Signature**		**Date**		**Time**	

Governor's Office Of Homeland Security & Emergency Preparedness

ORGANIZATION ASSIGNMENT LIST		Incident Number	15-005	ICS 203
Incident Name	Tulane National Primate Research Center	**Operational Period**	4/03/2015 0900hrs to 4/10/2015 0900hrs	
Unified Command		**Operations Section**		
UC/IC	RADM Scott Deitchman, MD (CDC)	Operations Chief	Miguel Cruz, PhD (CDC)	
UC/IC	Dir. Kevin Davis (GOHSEP)	Dep. Ops Chief	Collins Simoneaux (GOHSEP)	
UC/IC	Jimmy Guidry, MD (LA DHH)	**Branch 1 - Investigation Branch**		
UC/IC	Dir. Dexter Accardo (St Tammany OHSEP)	Branch Director	Robbin Weyant, PhD (CDC/OPHPR))	
Command Staff		S.A.P. Microbiologist	J.T. Paulley (CDC/OPHPR)	
Safety Officer	Dr. Don Sibley, PhD (Tulane)	S.A.P. Microbiologist	Lyla Lipscomb (CDC/OPHPR)	
PIO (JIC Primary Contacts)	* See PIO - JIC Information Below *	Medical Off. & SME	David Blaney (CDC/NCEZID)	
Liaison Officer	Darryl Delatte (GOHSEP)	EIS Officer	LCDR Leisha Nolen, MD (CDC/NCEZID)	
Agency Representative		EIS Officer	CDR Sherry Burrer, MD	
Tulane Primate Center	Dir. Mark Lackner, DVM	Industrial Hygenist	Joshua Harney (CDC/NIOSH)	
Tulane Primate Center	Mark Alise, PhD	Environmental Health Off.	LT James Gooch (CDC/NCEH)	
USDA/APHIS	Kenneth Angel, DVM	Public Health Scientist	LT Anna Kahn (CDC/OPHPR)	
USDA/APHIS	Katie Portacci, DVM	**Branch 2 - Remediation/Response Branch**		
USDA/APHIS	Lily Rai, DVM	Branch Director	Bill Rhotenberry (EPA)	
USDA/APHIS	Vicki Guilfor, DVM	Dep. Branch Dir.	Mike McAteer (EPA)	
USDA/APHIS	Freeda Isaac, DVM	Tech Specialist - SSC	Mike Nalipinski (EPA)	
US EPA	John Martin			
LDAF	Brent Robbins, DVM	**Joint Information Center**		
LA DHH	Gary Balsamo, DVM	Tulane PIO	Mike Strecker	
LA DHH	Dr. Roseanne Pratts	Tulane PIO	Debbie Grant	
LDEQ	Mike Algero	CDC PIO (Lead)	Barbara Reynolds	
LDEQ	Jeff Dauzat	CDC PIO	Jason McDonald	
FBI	Roger Tomberlin	CDC PIO	Christian Scheel	
DHS	Phillip Constantin	CDC PIO	Bernadette Burdin	
Planning Section		US EPA PIO	Joseph Hubbard	
Planning Chief	Paul Reeb (St. Tammany Parish)	US EPA PIO	David Gray	
Dep. Planning Chief		GOHSEP	Mike Steele	
Situation Unit		LA DHH	Olivia Watkins	
Resource Unit		LDAF	Veronica Mosgrove	
Documentation Unit	Vanessa Wall (St. Tammany Parish)	St. Tammany Parish	Ronnie Simpson	
Demobilization Unit				
Technical Specialists (Specialty and Name)		**Plans Section - Science Technical Working Group**		
*** Note - The Science Technical Working Group has been*		CDC - Working Group Lead	RADM Scott Deitchman, MD	
created under Plans Section. See adjacent list of members and		CDC - Human Blood Specialist	Dr. Henry Walke	
Objective 4 on the ICS-202.		CDC - Human Blood Specialist	Ermias Belay	
Logistics Section		CDC - Data Custodian	LT James Gooch	
Logistics Chief	Clarence Powe (St. Tammany Parish)	USDA - Research Leader	Dr. Alejandro Rooney	
Dep. Logistics Chief		EPA - Director, CMAD	Erica Canzler	
Service Branch		EPA - SSC	Mike Nalipinski	
Service Branch Dir.		DHH - State Health Officer	Dr. J. Guidry / Dr. R. Prats	
Communications Unit		DHH - State Epidemiologist	Dr. R. Ratard / Theresa Sokol	
IT Unit		DHH - State Lab	Dr. Stephen Martin	
Medical Unit		DHH - Regional MD	Dr. G. LaGarde / Dr. T. Davis	
Support Branch		LDAF - Asst. State Veterinarian	Dr. Diane Stacy, DVM	
Support Branch Dir.		LDEQ - ES Manager	Mike Algero	
Supply Unit		LDWF- State Wildlife Veterinarian	Dr. James Lacour, DVM	
Facilities Unit		Tulane - Dir. TNPRC	Dr. Andrew Lackner, DVM	
Finance Section		Tulane - Dir. Office of BioSafety	Dr. Don Sibley, PhD	
Finance Chief				

Prepared by	C.R.Simoneaux	**Signature**		**Date**		**Time**	
Approved by		**Signature**		**Date**		**Time**	

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INCIDENT TELEPHONE/E-MAIL CONTACT LIST		Incident Number	15-005	ICS 205T
Incident Name	Tulane National Primate Research Center	Operational Period	4/03/2015 0900hrs to 4/10/2015 0900hrs	

Telephone/Email List			
Agency/Position	Name	Cell Phone	Email
** Note - See Last Page for PIO/JIC and Public Inquiry Phone Numbers **			
Tulane National Primate Research Center			
Tulane - Director	Andrew Lackner, DVM	985-966-6402	alackner@tulane.edu
Tulane - COO	Mark Alise, PhD	985-871-6200	malise@tulane.edu
Tulane - Dir. Office of Biosafety	Dr. Don Sibley, PhD	504-988-0300	dsibley@tulane.edu
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Tulane - PIO	Debbie Grant	504-578-4071	dgrant@tulane.edu
St. Tammany Parish Contacts			
St Tammany Parish - President	Patricia Brister	985-898-2362	pbrister@stpgov.org
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St Tammany Parish - OHSEP	Vanessa Wall	985-898-2359	vanessa@stpgov.org
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GOHSEP - Logistics Chief	Michael Hamilton	225-335-1226	michael.hamilton@la.gov
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GOHSEP - PIO	Mike Steele	225-788-0095	mike.steele@la.gov
LA DHH - Secretary	Kathy Kliebert	225-342-9509	kathy.kliebert@la.gov
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LA DHH - Region 9 Medical Dir.	Dr. Gina Lagarde	985-285-7257	gina.lagarde@la.gov

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LDAF - Commissioner	Mike Strain, DVM	225-922-1233	mike.strain@ldaf.state.la.us
LDAF - State Veterinarian	Brent Robbins, DVM	985-264-4142	brobbins@ldaf.la.gov
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LDAF - Asst. Commissioner	John Walther	985-438-6111	john_walther@ldaf.state.la.us
LDAF - PIO	Veronica Mosgrove	225-229-7147	vmosgrove@ldaf.la.gov
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LDEQ - ESS	Jeff Dauzat	504-736-7714	jeff.dauzat@la.gov
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LDWF - Chief of Fisheries	Scott Longman	225-763-3513	slongman@wlf.lag.gov
LDWF - Colonel of Enforcement Div	Joey Broussard	225-765-2988	jbroussard@wlf.la.gov
LDWF - State Wildlife Veterinarian	James LaCour, DVM	225-907-9491	jmlacour@wlf.la.gov
LDWF - Asst. State Wildlife Vet.	Dr. Rusty Berry	318-487-5885	rberry@wlf.la.gov
Federal Agency Contacts			
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US CDC - Select Agent Program	Lyla Lipscomb	404-823-4910	vsw9@cdc.gov
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US CDC - EIS Officer	CDR Sherry Burrer, MD	614-507-9385	hhf8@cdc.gov
US CDC - Environ. Health Officer	LT James Gooch	404-444-3931	jsx5@cdc.gov
US CDC - NIOSH Industrial Hygenis	Joshua Harney	513-841-4202	vzh5@cdc.gov
US CDC - Human Blood Specialist	Dr. Henry Walke	404-452-9624	hfw3@cdc.gov
US CDC - Human Blood Specialist	Ermia Belay	404-444-3931	ebb8@cdc.gov
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US CDC - PIO	Jason McDonald	404-387-3660	gnf0@cdc.gov
US CDC - PIO	Bernadette Burden	404-213-6874	btb8@cdc.gov
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US EPA	Mike McAteer	214-354-9371	mcateer.mike@epa.gov
US EPA - OSC	John Martin	214-789-1994	martin.john@epa.gov
US EPA - Director, CMAD	Erica Canzler	202-564-2359	canzler.erica@epa.gov
US EPA - SSC	Mike Nalipinski	617-680-5469	nalipinski.mike@epa.gov

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US EPA - START	Robert Sherman	225-573-9785	robert.sherman@westonsolutions.com
US EPA - PIO	Joseph Hubbard	214-665-2200	hubbard.joseph@epa.gov
US EPA - PIO	David Gray	214-789-2619	gray.david@epa.gov
USDA - Asst Dist Director, Dist 4	Kenneth Angel, DVM	601-214-0722	kenneth.l.angel@aphis.usda.gov
USDA - Risk Analysis Proj. Mgr.	Katie Portacci, DVM	970-494-1741	katie.portacci@aphis.usda.gov
USDA -	Victoria Guilfoil, DVM	405-623-6505	victoria.y.guilfoil@aphis.usda.gov
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USDA - Dir. Ag Select Agent Service	Freeda Isaac, DVM	240-328-8927	freeda.e.isaac@aphis.usda.gov
USDA - PIO	Joelle Hayden	301-851-4040	joelle.r.hayden@aphis.usda.gov
FBI - WMD Program Coordinator	Roger Tomberlin	504-816-3035	roger.tomberlin@ic.fbi.gov
DHS - Protective Security Advisor	Phillip Constantin	504-202-1081	phillip.constantin@hq.dhs.gov
Maryland State Agency Contacts			
MEMA - Executive Director	Clay Stamp	410-517-3625	clay.stamp@maryland.gov
MD Dept. of Ag - Asst. State Vet.	Nancy Jo Chapman, DVM	410-841-5810	njo.chapman@maryland.gov
INCIDENT PIOs - JIC Contacts			
Tulane - PIO (Public Inquiries)	Mike Strecker	504-512-1347	mstreck@tulane.edu
Tulane - PIO	Debbie Grant	504-578-4071	dgrant@tulane.edu
US CDC - PIO (CDC Lead)	Barbara Reynolds, PhD	404-918-1586	bsr0@cdc.gov
US CDC - PIO	Christian Scheel	404-488-0583	bjn2@cdc.gov
US CDC - PIO (Public Inquiries)	Jason McDonald	404-387-3660	gnf0@cdc.gov
US CDC - PIO	Bernadette Burden	404-213-6874	btb8@cdc.gov
US EPA - PIO	Joseph Hubbard	214-665-2200	hubbard.joseph@epa.gov
US EPA - PIO	David Gray	214-789-2619	gray.david@epa.gov
USDA - PIO	Joelle Hayden	301-851-4040	joelle.r.hayden@aphis.usda.gov
GOHSEP - PIO	Mike Steele	225-788-0095	mike.steele@la.gov
LA DHH - PIO	Olivia Watkins	225-610-6660	olivia.watkins@la.gov
LDAF - PIO	Veronica Mosgrove	225-229-7147	vmosgrove@ldaf.la.gov
St Tammany Parish - PIO	Ronnie Simpson	504-905-7841	rsimpson@stpgov.org
**** Public Information Numbers - Only the Below Information is Releasable to the Public ****			
Questions regarding the investigation or remediation activities are directed to CDC (Jason McDonald) at 404-387-3660			
Questions regarding the TNPRC facility are directed to Tulane (Mike Strecker) at 504-512-1347			
All other questions are directed to GOHSEP (Mike Steele) at Mike.Steele@La.gov			

Prepared by	C.R.Simoneaux	**Signature**		**Date/Time**		

Tulane National Primate Research Center
18703 3 Rivers Road, Covington, LA 70433

Personal Protective Equipment and Other Work Practice Requirements for All Employees Responding to the Tulane National Primate Research Center *Burkholderia pseudomallei* Response.

February 11, 2015

Purpose and Scope

- This document is intended for the protection of federal, state, and local government employees as well as Tulane employees involved in the current *Burkholderia pseudomallei* investigation at the Tulane National Primate Research Center (TNPRC). Response workers are involved in the *Burkholderia pseudomallei* response on-site for the primary purpose of observation of response activities. Additional work activities may include: inspecting research facilities to include laboratories; interviewing employees, and reviewing TNPRC plans, SOPs, and work practices for handling select agents. Additional activities identified as part of the response may require another risk assessment that could result in the need for additional protection.
- This guidance outlines required personal protective equipment (PPE) and other work precautions that must be followed to protect employee health and safety and ensure biocontainment within the facility.
- EPA and their contractors are working under their own health and safety plan that is consistent with the requirements outlined in this document.

General Guidance

- Limited access. Access to the TNPRC, and especially the laboratories and animal colony, should be done with specific response or investigation duties.
- Basic entry requirements. For access to the laboratories and colony, individuals must be appropriately trained, fit tested (if a tight-fitting respirator), and have appropriate medical clearance for use of a respirator. Access to the laboratories and colony areas is limited to only individuals with a negative PPD TB test within the past 6 months.
- Open wounds. Any open wounds must be covered prior to entrance to the facility.
- All donning and doffing of PPE must occur with a "buddy" in order to ensure proper procedures are followed.
- Biocontainment practices should also be followed to avoid transiting potentially contaminated dust and soil from one animal cage to another.

Administrative Areas

- No specific PPE required, although individuals must cover any open wounds. TNPRC will be asked to provide foot baths for use by individuals who are returning from the primate grounds and associated areas who do not wear disposable foot coverings.

Laboratories

- CDC requested that TNPRC conduct cleaning and decontamination of any anterooms in which our participants will enter without PPE.
- Laboratory spaces: Follow TNPRC laboratory PPE requirements which include the use of respiratory protection, disposable gloves, gowns or coveralls, foot coverings, head coverings, and eye protection. PAPRs with disposable hoods equipped with high efficiency filters may be used in lieu of N-95 respirators. Double gloves must be worn in areas where select agent work has taken place but not been decontaminated.
- Hand hygiene must be performed after removal of PPE and upon exiting vivarium or animal colony rooms, necropsy rooms, and/or laboratory areas. Ensure that hands are thoroughly washed with soap and water; if soap and water are not readily available, use an alcohol-based hand rub. Follow with soap and water once available.

Non-Human Primate Colony- South Campus

- PPE described in this area is required for all staff observing activities related to the collection of animal serology and the investigation. All staff should maintain a safe distance of the animal enclosures whenever possible. Entrances and work within 10 feet of the animal enclosures will be kept to a minimum. TNPRC staff should be utilized as much as possible to work in this area under USDA observation and direction.
- Inquire with Tulane NPRC if there is a NHP "Bite, Scratch and Splash" kit readily available within premises in the possible event of an animal escaping its primary enclosure (e.g., caging).
- Recommended PPE includes:
    - Tyvek coveralls (with attached hood is preferred),
    - Boot covers,
    - Double gloves, and
    - N-95 filtering facepiece, face shield or goggles, and head cover,

      OR
    - Elastomeric respirator* equipped with high efficiency particulate air filter and head cover,

      OR
    - Powered air purifying respirator* equipped with high efficiency filter with disposable hood

*These higher levels of respiratory protection are recommended as an option because they offer protection of the mucous membranes, including the eyes and nose, and obviate the need for additional eye and face protection.

- Tulane will identify a dedicated sally port entrance to serve as the federal entrance/exit point. PPE must be donned and doffed in the area between the exterior gate and interior gate at the sally port.

- Boot covers must be removed and replaced with new boot covers before entering vehicles and between animal cages to avoid potential cross-contamination.

- All personnel on-site will follow these procedures:
    - The number of personnel entering the animal colony should be kept to an absolute minimum.
    - Entry to the animal colony will be through the sally port. PPE will be donned at this point prior to entry through the interior gate.
    - Donning and doffing will occur at the sally port with a buddy or observer to assist as needed.
    - Hand sanitizer will be available and used after doffing or assisting someone else in doffing.
    - Personnel will shower at the on-site facility only if direct contact with potentially infectious material (e.g., through a splash) or direct contact with an animal has occurred.
    - Personnel will minimize contact with any surfaces and remain as far from the primate cages as practicable while observing activities.
    - PPE will be collected in biohazard disposal bags and handled as hazardous material.
    - Personnel are encouraged to shower and shampoo as soon as possible after leaving the facility.
    - Any breaches in PPE (e.g., tear on glove) or any potential exposure while not wearing the appropriate PPE should be reported to the immediate supervisor or chain of command. Respective agency occupational health programs should also be notified.

- Hand hygiene must be performed after removal of PPE and upon exiting the colony area. Ensure that hands are thoroughly washed with soap and water; if soap and water are not readily available, use an alcohol-based hand rub. Follow with soap and water once available.

Surveillance for Occupational Infections

The signs and symptoms of melioidosis vary depending on the type of infection and can easily be mistaken for other diseases such as pneumonia or tuberculosis. The incubation period ranges from one day to many years, with the vast majority of cases presenting within 21 days of exposure.

- For wound infections, signs and symptoms include localized pain or swelling.

- For pulmonary infections, signs and symptoms include cough, headache, fever, chest pain, anorexia (lack of appetite), and general muscle soreness.

- For bloodstream infections, signs and symptoms include high fever, headache, respiratory distress, disorientation, abdominal discomfort, and muscle tenderness.

- For infections that have spread throughout the body, signs and symptoms include fever, weight loss, stomach or chest pain, muscle or joint pain, headache, and encephalitis or seizures.

Self-monitoring:

Employees should do active daily self-monitoring for the presence of symptoms of melioidosis and fever for 21 days after the last day on site at TNPRC. Any fever above normal must be reported to the immediate supervisor or chain of command, and respective agency health and safety staff. Collection of blood for employees prior to visiting the site is recommended to serve as baseline samples should they be needed. Banked samples will not be analyzed at the time of collection, rather decisions on the need to analyze bank sera will be determined by the Unified Command.

Dan A. Lebby 2/12/2015
Safety Officer BP Response

Cleaning and Disinfecting Vehicles

All vehicles (for example, cars, livestock carriers, feed trucks, milk trucks, and carcass transporters) and heavy machinery (for example, excavators, backhoes, and bulldozers) that have been used on the Tulane National Primate Research Center, South Campus as part of the B pseudomallei response must undergo proper C&D processes before departing the premises because of the potential to transport pathogens across premises.

General Procedures

1. Don PPE as approved in the safety plan.
2. Remove the following items and set these items at the appropriate steps for separate C&D
a. Equipment in the truck bed or trailer, or in the vehicle's trunk.
b. Fixtures and fittings.
c. items should be returned to the vehicle after cleaning and/or disinfections are complete.

Cleaning Procedures

1. Dispose of soiled bedding and refuse and debris cleaned from the vehicle in appropriate biohazard containers.
2. Use shovels, manure forks, brushes, low-pressure sprayers, or mechanical scrapers to remove all visible organic material from the exterior of the vehicle. Remove any deposits of mud and straw from the wheels, wheel wells, tires, mudguards, and exposed chassis of the vehicle. It is essential that appropriate PPE be worn, especially when zoonotic disease agents are involved.

Washing Procedures

1.The power wash at Z Building will be used to clean the exterior of vehicles of all soil accumulation as well as to clean soiled tools and implements used in the completion of tasks associated with the B pseudomallei response mission in order to allow effective decontamination with liquid disinfectant. Any deposits of mud and straw should be removed from the wheels, wheel wells, tires, mudguards, and exposed chassis of the vehicle.
2. If interior of the vehicle has been contaminated with soil or debris remove the soil and debris and if necessary remove the floor mats or other items for washing with the power washer.
3. Following the washing procedure the vehicle, tools and other items washed will be rinsed with water and allow 5–10 minutes for residual rinse water to drip off.
4. Remove PPE soiled during the washing process and replace with clean PPE.

Exterior Disinfection

1. At the sally port the exterior of the vehicle and any items washed in the previous step will be sprayed with a 1% Virkon S® solution as a disinfectant. A garden type pump sprayer or equivalent can be used for application.
2. For vehicles and trailers, apply the disinfectant to the exterior of the vehicle, including the bodywork and wheels.
3. For trailers, apply the disinfectant to the outside of the trailer and the underside of the vehicle.

4. Spray all areas, including the wheels, wheel wells, tires, mudguards, and exposed chassis of the vehicle with disinfectant.
5. Allow 10 minutes of wet disinfectant contact time with the vehicle or trailer.

Interior Cleaning and Disinfection

Interior disinfection of the vehicle is necessary if the driver or passengers leave the cab. If so, all surfaces on the interior of the cab will need to be disinfected.

1. Remove all non-fixed items from the vehicle to be disinfected.
2. Disinfect the interior surfaces of the vehicle with 1% Virkon S® solution and allow 10 minutes contact time before personnel re-enter the vehicle. After the contact time is complete, the surfaces may be rinsed or wiped down with clean water.
3. Replace all items removed for disinfection and dispose of PPE.
4. Re- enter vehicle and drive out of the sally port.

Don Sibley, Ph.D., Safety Officer 02/12/2015